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(21) International Application Number: PCT/US98/13859 (22) International Filing Date: 2 July 1998 (02.07.98) (30) Priority Data: <table border="0" style="width: 100%;"> <tr> <td style="width: 30%;">60/051,704</td> <td style="width: 40%;">3 July 1997 (03.07.97)</td> <td style="width: 30%;">US</td> </tr> <tr> <td>08/888,088</td> <td>3 July 1997 (03.07.97)</td> <td>US</td> </tr> <tr> <td>60/085,983</td> <td>19 May 1998 (19.05.98)</td> <td>US</td> </tr> <tr> <td>09/081,338</td> <td>19 May 1998 (19.05.98)</td> <td>US</td> </tr> <tr> <td>60/089,899</td> <td>17 June 1998 (17.06.98)</td> <td>US</td> </tr> <tr> <td>09/099,005</td> <td>17 June 1998 (17.06.98)</td> <td>US</td> </tr> </table> (71) Applicant: ZYMOGENETICS, INC. [US/US]; 1201 Eastlake Avenue East, Seattle, WA 98102 (US). (72) Inventors: SHEPPARD, Paul, O.; 20717 N.E. 2nd Street, Redmond, WA 98053 (US). JELINEK, Laura, J.; 1124 N.E. 147th, Seattle, WA 98155 (US). JASPERS, Stephen, R.; 5829 150th Place S.W., Edmonds, WA 98026 (US). WHITMORE, Theodore, E.; 6916 152nd Avenue N.E., Redmond, WA 98052 (US). (74) Agent: LUNN, Paul, G.; ZymoGenetics, Inc., 1201 Eastlake Avenue East, Seattle, WA 98102 (US).		60/051,704	3 July 1997 (03.07.97)	US	08/888,088	3 July 1997 (03.07.97)	US	60/085,983	19 May 1998 (19.05.98)	US	09/081,338	19 May 1998 (19.05.98)	US	60/089,899	17 June 1998 (17.06.98)	US	09/099,005	17 June 1998 (17.06.98)	US	(81) Designated States: AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, CA, CH, CN, CU, CZ, DE, DK, EE, ES, FI, GB, GE, GH, GM, GW, HR, HU, ID, IL, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MD, MG, MK, MN, MW, MX, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TR, TT, UA, UG, UZ, VN, YU, ZW, ARIPO patent (GH, GM, KE, LS, MW, SD, SZ, UG, ZW), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, ML, MR, NE, SN, TD, TG). Published <i>With international search report.</i> <i>Before the expiration of the time limit for amending the claims and to be republished in the event of the receipt of amendments.</i>
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(54) Title: MAMMALIAN SECRETORY PEPTIDE-9																				
(57) Abstract <p>Novel mammalian secretory polypeptides, polynucleotides encoding the polypeptides (called Zsig9), and related compositions and methods including antibodies and antiidiotypic antibodies. Overexpression of these proteins is indicative of the presence of cancer. Antibodies and antisense nucleotides can be used therapeutically to treat the disease. Furthermore, antibodies to Zsig9 and nucleotide primers and probes can be used to diagnose the presence of tumors.</p>																				

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5 MAMMALIAN SECRETORY PEPTIDE - 9

BACKGROUND OF THE INVENTION

10 Proliferation and differentiation of cells of multicellular organisms are controlled by hormones and polypeptide growth factors. These diffusable molecules allow cells to communicate with each other and act in concert to form cells and organs, and to repair and regenerate damaged tissue. Examples of hormones and
15 growth factors include the steroid hormones (e.g. estrogen, testosterone), parathyroid hormone, follicle stimulating hormone, the interleukins, platelet derived growth factor (PDGF), epidermal growth factor (EGF), granulocyte-macrophage colony stimulating factor (GM-CSF),
20 erythropoietin (EPO) and calcitonin.

Hormones and growth factors influence cellular metabolism by binding to proteins. Proteins may be integral membrane proteins that are linked to signaling
25 pathways within the cell, such as second messenger systems. Other classes of proteins are soluble molecules, such as the transcription factors.

Thus, there is a continuing need to discover new
30 hormones, growth factors and the like.

SUMMARY OF THE INVENTION

The present invention addresses this need by
35 providing a novel polypeptide and related compositions and methods. Within one aspect, the present invention provides

an isolated polynucleotide encoding a mammalian polypeptide termed secretory peptide - 9, hereinafter referred to as Zsig9. The mature human Zsig9 polypeptide is comprised of a sequence of amino acids approximately 64 amino acids long. Amino acid residue 21 of SEQ ID NO: 2, an arginine, is the initial amino acid of the mature polypeptide. Thus, it is believed that amino residues 1-20 comprise a signal sequence, and the mature Zsig9 polypeptide is represented by the amino acid sequence comprised of residues 21-84. The mature Zsig9 polypeptide is further represented by SEQ ID NO: 3. Within an additional embodiment, the polypeptide further comprises an affinity tag. Within a further embodiment, the polynucleotide is DNA.

15

Alternative forms of Zsig9 are defined by SEQ ID NOs: 4, 5, and 6. SEQ ID NO: 4 defines a processed form of Zsig9 in which the protein contains amino acid residues 23 - 84 of SEQ ID NO: 2.

20

SEQ ID NO: 5 represents another form of Zsig9 containing amino acid residues 23, a serine, to and including amino acid 47, a proline of SEQ ID NO: 2.

25

SEQ ID NO: 6 defines another processed form of Zsig9 contain amino acid residues 50, a threonine, to and including amino acid 84 of SEQ ID NO: 2.

SEQ ID NO: 16 and 17 represent another variant of Zsig9 and SEQ ID NO: 20 represents the mature sequence absent the first 20 amino acid residues, the signal sequence, of SEQ ID NO: 17.

SEQ ID NO: 18 and 19 represent the mouse ortholog of Zsig9; and SEQ ID NO: 21 depicts the mature amino acid

35

sequence absent the first 20 amino acid residues, the signal sequence, of SEQ ID NO: 19.

Within a second aspect of the invention there is
5 provided an expression vector comprising (a) a transcription promoter; (b) a DNA segment encoding a Zsig9 polypeptide as defined by SEQ ID NOs: 2-6, 17, 20 19 and 21 or a polypeptide 90% identical to said polypeptides, and (c) a transcription terminator, wherein
10 the promoter, DNA segment, and terminator are operably linked.

Within a third aspect of the invention there is provided a cultured eukaryotic cell into which has been
15 introduced an expression vector as disclosed above, wherein said cell expresses a protein polypeptide encoded by the DNA segment.

Within a further aspect of the invention there is
20 provided a chimeric polypeptide consisting essentially of a first portion and a second portion joined by a peptide bond. The first portion of the chimeric polypeptide consists essentially of (a) a Zsig9 polypeptide as shown in SEQ ID NOs: 2-6, 17, 20 19 and 21 (b) allelic variants
25 of SEQ ID NOs: 2-6, 17, 20 19 and 21; and (c) protein polypeptides that are at least 90% identical to (a) or (b). The second portion of the chimeric polypeptide consists essentially of another polypeptide such as an affinity tag. Within one embodiment the affinity tag is
30 an immunoglobulin F_C polypeptide. The invention also provides expression vectors encoding the chimeric polypeptides and host cells transfected to produce the chimeric polypeptides.

35 Within an additional aspect of the invention there is provided an antibody that specifically binds to a Zsig9

polypeptide as disclosed above, and also an anti-idiotypic antibody which neutralizes the antibody to a Zsig9 polypeptide.

5 An additional embodiment of the present invention relates to a peptide or polypeptide which has the amino acid sequence of an epitope-bearing portion of a Zsig9 polypeptide having an amino acid sequence described above. Peptides or polypeptides having the amino acid sequence of
10 an epitope-bearing portion of a Zsig9 polypeptide of the present invention include portions of such polypeptides with at least nine, preferably at least 15 and more preferably at least 30 to 50 amino acids, although epitope-bearing polypeptides of any length up to and
15 including the entire amino acid sequence of a polypeptide of the present invention described above are also included in the present invention. An example of such a polypeptide is represented by SEQ ID NO:24.

20 Also claimed are antibodies which bind specifically to the above-defined Zsig9 polypeptides, methods for making said antibodies, and anti-idiotypic antibodies of said Zsig9-binding antibodies.

25 These and other aspects of the invention will become evident upon reference to the following detailed description.

30 DETAILED DESCRIPTION OF THE INVENTION

The teachings of all of the references cited herein are incorporated in their entirety herein by reference.

35 The term "ortholog" (or "species homolog") denotes a polypeptide or protein obtained from one species that has

homology to an analogous polypeptide or protein from a different species.

The term "paralog" denotes a polypeptide or protein
5 obtained from a given species that has homology to a distinct polypeptide or protein from that same species.

The term "allelic variant" denotes any of two or more alternative forms of a gene occupying the same chromosomal
10 locus. Allelic variation arises naturally through mutation, and may result in phenotypic polymorphism within populations. Gene mutations can be silent (no change in the encoded polypeptide) or may encode polypeptides having altered amino acid sequence. The term allelic variant is
15 also used herein to denote a protein encoded by an allelic variant of a gene.

The term "expression vector" denotes a DNA molecule, linear or circular, that comprises a segment encoding a
20 polypeptide of interest operably linked to additional segments that provide for its transcription. Such additional segments include promoter and terminator sequences, and may optionally include one or more origins of replication, one or more selectable markers, an
25 enhancer, a polyadenylation signal and the like. Expression vectors are generally derived from plasmid or viral DNA, or may contain elements of both.

The term "isolated", when applied to a
30 polynucleotide, denotes that the polynucleotide has been removed from its natural genetic milieu and is thus free of other extraneous or unwanted coding sequences, and is in a form suitable for use within genetically engineered protein production systems. Such isolated molecules are
35 those that are separated from their natural environment and include cDNA and genomic clones. Isolated DNA

molecules of the present invention are free of other genes with which they are ordinarily associated, but may include naturally occurring 5' and 3' untranslated regions such as promoters and terminators. The identification of
5 associated regions will be evident to one of ordinary skill in the art, Dynan and Tijan, *Nature* 316:774-78 (1985). When applied to a protein, the term "isolated" indicates that the protein is found in a condition other than its native environment, such as apart from blood and
10 animal tissue. In a preferred form, the isolated protein is substantially free of other proteins, particularly other proteins of animal origin. It is preferred to provide the protein in a highly purified form, i.e., greater than 95% pure, more preferably greater than 99%
15 pure.

The term "operably linked", when referring to DNA segments, denotes that the segments are arranged so that they function in concert for their intended purposes, e.g.
20 transcription initiates in the promoter and proceeds through the coding segment to the terminator.

The term "polynucleotide" is a single- or double-stranded polymer of deoxyribonucleotide or ribonucleotide
25 bases read from the 5' to the 3' end. Polynucleotides include RNA and DNA, and may be isolated from natural sources, synthesized *in vitro*, or prepared from a combination of natural and synthetic molecules.

30 The term "complements of polynucleotide molecules" denotes polynucleotide molecules having a complementary base sequence and reverse orientation as compared to a reference sequence.

35 The term "degenerate nucleotide sequence" denotes a sequence of nucleotides that includes one or more

degenerate codons (as compared to a reference polynucleotide molecule that encodes a polypeptide). Degenerate codons contain different triplets of nucleotides, but encode the same amino acid residue (i.e., GAU and GAC triplets each encode Asp).

The term "promoter" denotes a portion of a gene containing DNA sequences that provide for the binding of RNA polymerase and initiation of transcription. Promoter sequences are commonly, but not always, found in the 5' non-coding regions of genes.

The term "secretory signal sequence" denotes a DNA sequence that encodes a polypeptide (a "secretory peptide") that, as a component of a larger polypeptide, directs the larger polypeptide through a secretory pathway of a cell in which it is synthesized. The larger peptide is commonly cleaved to remove the secretory peptide during transit through the secretory pathway.

The term "receptor" denotes a cell-associated protein that binds to a bioactive molecule (i.e., a ligand) and mediates the effect of the ligand on the cell. Membrane-bound receptors are characterized by a multi-domain structure comprising an extracellular ligand-binding domain and an intracellular effector domain that is typically involved in signal transduction. Binding of ligand to receptor results in a conformational change in the receptor that causes an interaction between the effector domain and other molecule(s) in the cell. This interaction in turn leads to an alteration in the metabolism of the cell. Metabolic events that are linked to receptor-ligand interactions include gene transcription, phosphorylation, dephosphorylation, increases in cyclic AMP production, mobilization of cellular calcium, mobilization of membrane lipids, cell

adhesion, hydrolysis of inositol lipids and hydrolysis of phospholipids. Most nuclear receptors also exhibit a multi-domain structure, including an amino-terminal, transactivating domain, a DNA binding domain and a ligand binding domain. In general, receptors can be membrane bound, cytosolic or nuclear; monomeric (e.g., thyroid stimulating hormone receptor, beta-adrenergic receptor) or multimeric (e.g., PDGF receptor, growth hormone receptor, IL-3 receptor, GM-CSF receptor, G-CSF receptor, erythropoietin receptor and IL-6 receptor).

The term "complement/anti-complement pair" denotes non-identical moieties that form a non-covalently associated, stable pair under appropriate conditions. For instance, biotin and avidin (or streptavidin) are prototypical members of a complement/anti-complement pair. Other exemplary complement/anti-complement pairs include receptor/ligand pairs, antibody/antigen (or hapten or epitope) pairs, sense/antisense polynucleotide pairs, and the like. Where subsequent dissociation of the complement/anti-complement pair is desirable, the complement/anti-complement pair preferably has a binding affinity of $<10^9 \text{ M}^{-1}$.

A "soluble protein" is a protein polypeptide that is not bound to a cell membrane.

Within preferred embodiments of the invention the isolated polynucleotides will hybridize to similar sized regions of polynucleotide defined by SEQ ID NO:1, or a sequence complementary thereto, under stringent conditions. In general, stringent conditions are selected to be about 5°C lower than the thermal melting point (T_m) for the specific sequence at a defined ionic strength and pH. The T_m is the temperature (under defined ionic

strength and pH) at which 50% of the target sequence hybridizes to a perfectly matched probe. Typical stringent conditions are those in which the salt concentration is about 0.02 M or less at pH 7 and the temperature is at least about 60°C. As previously noted, the isolated polynucleotides of the present invention include DNA and RNA. Methods for isolating DNA and RNA are well known in the art. Total RNA can be prepared using guanidine HCl extraction followed by isolation by centrifugation in a CsCl gradient, Chirgwin et al., *Biochemistry* 18:52-94 (1979). Poly (A)⁺ RNA is prepared from total RNA using the method of Aviv and Leder, *Proc. Natl. Acad. Sci. USA* 69:1408-1412 (1972). Complementary DNA (cDNA) is prepared from poly(A)⁺ RNA using known methods. Polynucleotides encoding Zsig9 polypeptides are then identified and isolated by, for example, hybridization or PCR.

The present invention further provides counterpart polypeptides and polynucleotides from other species (orthologs or paralogs). Of particular interest are Zsig9 polypeptides from other mammalian species, including murine, rat, porcine, ovine, bovine, canine, feline, equine and other primate proteins. Species homologs of the human proteins can be cloned using information and compositions provided by the present invention in combination with conventional cloning techniques. For example, a cDNA can be cloned using mRNA obtained from a tissue or cell type that expresses the protein. Suitable sources of mRNA can be identified by probing Northern blots with probes designed from the sequences disclosed herein. A library is then prepared from mRNA of a positive tissue or cell line. A Zsig9-encoding cDNA can then be isolated by a variety of methods, such as by probing with a complete or partial human cDNA or with one

or more sets of degenerate probes based on the disclosed sequences. A cDNA can also be cloned using the polymerase chain reaction, or PCR (Mullis, U.S. Patent 4,683,202), using primers designed from the sequences disclosed
5 herein. Within an additional method, the cDNA library can be used to transform or transfect host cells, and expression of the cDNA of interest can be detected with an antibody to Zsig9. Similar techniques can also be applied to the isolation of genomic clones.

10

Those skilled in the art will recognize that the sequences disclosed in SEQ ID NOs:1 and 2, SEQ ID NOs:16 and 17, SEQ ID NOs. 18 and 19, represent a specific alleles of the human Zsig9 gene and polypeptide, and that
15 allelic variation and alternative splicing are expected to occur. Allelic variants can be cloned by probing cDNA or genomic libraries from different individuals according to standard procedures. Allelic variants of the DNA sequence shown in SEQ ID NO: 1, including those containing silent
20 mutations and those in which mutations result in amino acid sequence changes, are within the scope of the present invention, as are proteins which are allelic variants of SEQ ID NOs: 3, 4, 5, and 6.

25 Additionally, the polynucleotides of the present invention can be synthesized using DNA synthesizers. Currently the method of choice is the phosphoramidite method. Each complementary strand of a double stranded DNA of gene or a gene fragment is made separately. The
30 production of short DNA fragments (60 to 80 bp) can be accomplished by synthesizing the complementary strands and then annealing them. For the production of longer DNA molecules the coupling efficiency of each cycle during chemical DNA synthesis is seldom 100%. To overcome this
35 problem, synthetic genes (double-stranded) are assembled

in modular form from single-stranded fragments that are from 20 to 100 nucleotides in length.

One method for building a synthetic gene requires the initial production of a set of overlapping, complementary oligonucleotides, each of which is between 20 to 60 nucleotides long. The sequences of the strands are planned so that, after annealing, the two end segments of the gene are aligned to give blunt ends. Each internal section of the gene has complementary 3' and 5' terminal extensions that are designed to base pair precisely with an adjacent section. Thus, after the gene is assembled, the only remaining requirement to complete the process is sealing the nicks along the backbones of the two strands with T4 DNA ligase. In addition to the protein coding sequence, synthetic genes can be designed with terminal sequences that facilitate insertion into a restriction endonuclease sites of a cloning vector and other sequences should also be added that contain signals for the proper initiation and termination of transcription and translation. See Glick, Bernard R. and Jack J. Pasternak, *Molecular Biotechnology, Principles & Applications of Recombinant DNA*, (ASM Press, Washington, D.C. 1994), Itakura, K. et al. Synthesis and use of synthetic oligonucleotides. *Annu. Rev. Biochem.* 53 : 323-356 (1984), and Climie, S. et al. Chemical synthesis of the thymidylate synthase gene. *Proc. Natl. Acad. Sci. USA* 87 :633-637 (1990).

Those skilled in the art will recognize that the sequences disclosed in SEQ ID NOS:1, 2, 3, 4, 5, and 6 represent a single allele of the human. Allelic variants of these sequences can be cloned by probing cDNA or genomic libraries from different individuals according to standard procedures.

The present invention further provides counterpart proteins and polynucleotides from other species ("species orthologs"). Of particular interest are Zsig9 polypeptides from other mammalian species, including murine, porcine, ovine, bovine, canine, feline, equine, and other primates. Species orthologs of the human Zsig9 protein can be cloned using information and compositions provided by the present invention in combination with conventional cloning techniques. For example, a cDNA can be cloned using mRNA obtained from a tissue or cell type that expresses the protein. Suitable sources of mRNA can be identified by probing Northern blots with probes designed from the sequences disclosed herein. A library is then prepared from mRNA of a positive tissue or cell line. A protein-encoding cDNA can then be isolated by a variety of methods, such as by probing with a complete or partial human or mouse cDNA or with one or more sets of degenerate probes based on the disclosed sequences. A cDNA can also be cloned using the polymerase chain reaction, or PCR (Mullis, U.S. Patent No. 4,683,202), using primers designed from the sequences disclosed herein. Within an additional method, the cDNA library can be used to transform or transfect host cells, and expression of the cDNA of interest can be detected with an antibody to the protein. Similar techniques can also be applied to the isolation of genomic clones. As used and claimed the language "an isolated polynucleotide which encodes a polypeptide, said polypeptide being defined by SEQ ID NO: 2-6, 17, 20 19 and 21" includes all allelic variants and species orthologs of the polypeptide of SEQ ID NOs: 2-6, 17, 20 19 and 21.

The present invention also provides isolated protein polypeptides that are substantially homologous to the polypeptides of SEQ ID NOs: 2, 3, 4, 5 or 6 and their species orthologs. By "isolated" is meant a protein or

polypeptide that is found in a condition other than its native environment, such as apart from blood and animal tissue. In a preferred form, the isolated polypeptide is substantially free of other polypeptides, particularly other polypeptides of animal origin. It is preferred to provide the polypeptides in a highly purified form, i.e. greater than 95% pure, more preferably greater than 99% pure. The term "substantially homologous" is used herein to denote polypeptides having 50%, preferably 60%, more preferably at least 80%, sequence identity to the sequence shown in SEQ ID NO:2, or its species orthologs. Such polypeptides will more preferably be at least 90% identical, and most preferably 95% or more identical to SEQ ID NO:3, or its species orthologs. Percent sequence identity is determined by conventional methods. See, for example, Altschul et al., *Bull. Math. Bio.* 48: 603-616 (1986) and Henikoff and Henikoff, *Proc. Natl. Acad. Sci. USA* 89:10915-10919 (1992). Briefly, two amino acid sequences are aligned to optimize the alignment scores using a gap opening penalty of 10, a gap extension penalty of 1, and the "blossom 62" scoring matrix of Henikoff and Henikoff (ibid.) as shown in Table 2 (amino acids are indicated by the standard one-letter codes). The percent identity is then calculated as:

$$\frac{\text{Total number of identical matches}}{[\text{length of the longer sequence plus the number of gaps introduced into the longer sequence in order to align the two sequences}]} \times 100$$

Table 1

	A	R	N	D	C	Q	E	G	H	I	L	K	M	F	P	S	T	W	Y	V
5	A 4																			
	R -1	5																		
	N -2	0	6																	
	D -2	-2	1	6																
	C 0	-3	-3	-3	9															
	Q -1	1	0	0	-3	5														
10	E -1	0	0	2	-4	2	5													
	G 0	-2	0	-1	-3	-2	-2	6												
	H -2	0	1	-1	-3	0	0	-2	8											
	I -1	-3	-3	-3	-1	-3	-3	-4	-3	4										
	L -1	-2	-3	-4	-1	-2	-3	-4	-3	2	4									
15	K -1	2	0	-1	-3	1	1	-2	-1	-3	-2	5								
	M -1	-1	-2	-3	-1	0	-2	-3	-2	1	2	-1	5							
	F -2	-3	-3	-3	-2	-3	-3	-3	-1	0	0	-3	0	6						
	P -1	-2	-2	-1	-3	-1	-1	-2	-2	-3	-3	-1	-2	-4	7					
	S 1	-1	1	0	-1	0	0	0	-1	-2	-2	0	-1	-2	-1	4				
20	T 0	-1	0	-1	-1	-1	-1	-2	-2	-1	-1	-1	-1	-2	-1	1	5			
	W -3	-3	-4	-4	-2	-2	-3	-2	-2	-3	-2	-3	-1	1	-4	-3	-2	11		
	Y -2	-2	-2	-3	-2	-1	-2	-3	2	-1	-1	-2	-1	3	-3	-2	-2	2	7	
	V 0	-3	-3	-3	-1	-2	-2	-3	-3	3	1	-2	1	-1	-2	-2	0	-3	-1	4

Sequence identity of polynucleotide molecules is determined by similar methods using a ratio as disclosed above.

5 Substantially homologous proteins and polypeptides are characterized as having one or more amino acid substitutions, deletions or additions. These changes are preferably of a minor nature, that is conservative amino acid substitutions (see Table 3) and other substitutions
10 that do not significantly affect the folding or activity of the protein or polypeptide; small deletions, typically of one to about 30 amino acids; and small amino- or carboxyl-terminal extensions, such as an amino-terminal methionine residue, a small linker peptide of up to about
15 20-25 residues, or a small extension that facilitates purification (an affinity tag), such as a poly-histidine tract, protein A, Nilsson et al., *EMBO J.* 4:1075, (1985); Nilsson et al., *Methods Enzymol.* 198:3, (1991), glutathione S transferase, Smith and Johnson, *Gene* 67:31,
20 (1988), or other antigenic epitope or binding domain. See, in general Ford et al., *Protein Expression and Purification* 2: 95-107, (1991). DNAs encoding affinity tags are available from commercial suppliers (e.g., Pharmacia Biotech, Piscataway, NJ).

25

Table 2Conservative amino acid substitutions

30	Basic:	arginine
		lysine
		histidine
	Acidic:	glutamic acid
		aspartic acid
	Polar:	glutamine
		asparagine
35	Hydrophobic:	leucine

Table 2, continued

	isoleucine
	valine
Aromatic:	phenylalanine
5	tryptophan
	tyrosine
Small:	glycine
	alanine
	serine
10	threonine
	methionine

The proteins of the present invention can also comprise, in addition to the 20 standard amino acids, non-naturally occurring amino acid residues. Non-naturally occurring amino acids include, without limitation, trans-3-methylproline, 2,4-methanoproline, cis-4-hydroxyproline, trans-4-hydroxyproline, N-methyl-glycine, allo-threonine, methylthreonine, hydroxyethyl-cysteine, hydroxyethylhomocysteine, nitroglutamine, homoglutamine, pipecolic acid, tert-leucine, norvaline, 2-azaphenylalanine, 3-azaphenylalanine, 4-azaphenyl-alanine, 4-fluorophenylalanine, 4-hydroxyproline, 6-N-methyl lysine, 2-aminoisobutyric acid, isovaline and α -methyl serine. Several methods are known in the art for incorporating non-naturally occurring amino acid residues into proteins. For example, an *in vitro* system can be employed wherein nonsense mutations are suppressed using chemically aminoacylated suppressor tRNAs. Methods for synthesizing amino acids and aminoacylating tRNA are known in the art. Transcription and translation of plasmids containing nonsense mutations are carried out in a cell free system comprising an *E. coli* S30 extract and commercially available enzymes and other reagents.

Proteins are purified by chromatography. See, for example, Robertson et al., *J. Am. Chem. Soc.* 113:2722 (1991); Ellman et al., *Meth. Enzymol.* 202:301 (1991); Chung et al., *Science* 259:806-09 (1993); and Chung et al.,
5 *Proc. Natl. Acad. Sci. USA* 90:10145-49 (1993). In a second method, translation is carried out in *Xenopus* oocytes by microinjection of mutated mRNA and chemically aminoacylated suppressor tRNAs (Turcatti et al., *J. Biol. Chem.* 271:19991-98 (1996). Within a third method, *E. coli*
10 cells are cultured in the absence of a natural amino acid that is to be replaced (e.g., phenylalanine) and in the presence of the desired non-naturally occurring amino acid(s) (e.g., 2-azaphenylalanine, 3-azaphenylalanine, 4-azaphenylalanine, or 4-fluorophenylalanine). The non-
15 naturally occurring amino acid is incorporated into the protein in place of its natural counterpart. See, Koide et al., *Biochem.* 33:7470-76, 1994. Naturally occurring amino acid residues can be converted to non-naturally occurring species by in vitro chemical modification.
20 Chemical modification can be combined with site-directed mutagenesis to further expand the range of substitutions (Wynn and Richards, *Protein Sci.* 2:395-403 (1993).

A limited number of non-conservative amino acids,
25 amino acids that are not encoded by the genetic code, non-naturally occurring amino acids, and unnatural amino acids may be substituted for Zsig9* amino acid residues.
"Unnatural amino acids" have been modified after protein synthesis, and/or have a chemical structure in their side
30 chain(s) different from that of the standard amino acids. Unnatural amino acids can be chemically synthesized, or preferably, are commercially available, and include
pipecolic acid, thiazolidine carboxylic acid,

dehydroproline, 3- and 4-methylproline, and 3,3-dimethylproline.

Essential amino acids in the polypeptides of the present invention can be identified according to procedures known in the art, such as site-directed mutagenesis or alanine-scanning mutagenesis, Cunningham and Wells, *Science* 244, 1081-1085, (1989); Bass et al., *Proc. Natl. Acad. Sci. USA* 88:4498-4502, (1991). In the latter technique, single alanine mutations are introduced at every residue in the molecule, and the resultant mutant molecules are tested for biological activity (e.g., ligand binding and signal transduction) to identify amino acid residues that are critical to the activity of the molecule. Sites of ligand-protein interaction can also be determined by analysis of crystal structure as determined by such techniques as nuclear magnetic resonance, crystallography or photoaffinity labeling. See, for example, de Vos et al., *Science* 255:306-312, (1992); Smith et al., *J. Mol. Biol.* 224:899-904, (1992); Wlodaver et al., *FEBS Lett.* 309:59-64, (1992). The identities of essential amino acids can also be inferred from analysis of homologies with related proteins.

Multiple amino acid substitutions can be made and tested using known methods of mutagenesis and screening, such as those disclosed by Reidhaar-Olson and Sauer, *Science* 241:53-57, (1988) or Bowie and Sauer, *Proc. Natl. Acad. Sci. USA* 86:2152-2156, (1989). Briefly, these authors disclose methods for simultaneously randomizing two or more positions in a polypeptide, selecting for functional polypeptide, and then sequencing the mutagenized polypeptides to determine the spectrum of allowable substitutions at each position. Other methods that can be used include phage display, e.g., Lowman et

al., *Biochem.* 30:10832-10837, (1991); Ladner et al., U.S. Patent No. 5,223,409; Huse, WIPO Publication WO 92/06204, and region-directed mutagenesis, Derbyshire et al., *Gene* 46:145, (1986); Ner et al., *DNA* 7:127, (1988)

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Mutagenesis methods as disclosed above can be combined with high-throughput screening methods to detect activity of cloned, mutagenized proteins in host cells. Preferred assays in this regard include cell proliferation
10 assays and biosensor-based ligand-binding assays, which are described below. Mutagenized DNA molecules that encode active proteins or portions thereof (e.g., ligand-binding fragments) can be recovered from the host cells and rapidly sequenced using modern equipment. These methods
15 allow the rapid determination of the importance of individual amino acid residues in a polypeptide of interest, and can be applied to polypeptides of unknown structure.

20 Using the methods discussed above, one of ordinary skill in the art can prepare a variety of polypeptides that are substantially homologous to SEQ ID NO:2 or allelic variants thereof and retain the properties of the wild-type protein. As expressed and claimed herein the
25 language, "a polypeptide as defined by SEQ ID NO: 2" includes all allelic variants and species orthologs of the polypeptide.

The protein/polypeptides of the present invention,
30 including full-length proteins, protein fragments (e.g. ligand-binding fragments), and fusion polypeptides can be produced in genetically engineered host cells according to conventional techniques. Suitable host cells are those cell types that can be transformed or transfected with
35 exogenous DNA and grown in culture, and include bacteria, fungal cells, and cultured higher eukaryotic cells.

Eukaryotic cells, particularly cultured cells of multicellular organisms, are preferred. Techniques for manipulating cloned DNA molecules and introducing exogenous DNA into a variety of host cells are disclosed
5 by Sambrook et al., *Molecular Cloning: A Laboratory Manual*, 2nd ed., Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY, (1989), and Ausubel et al., *ibid.*

In general, a DNA sequence encoding a Zsig9
10 polypeptide is operably linked to other genetic elements required for its expression, generally including a transcription promoter and terminator, within an expression vector. The vector will also commonly contain one or more selectable markers and one or more origins of
15 replication, although those skilled in the art will recognize that within certain systems selectable markers may be provided on separate vectors, and replication of the exogenous DNA may be provided by integration into the host cell genome. Selection of promoters, terminators,
20 selectable markers, vectors and other elements is a matter of routine design within the level of ordinary skill in the art. Many such elements are described in the literature and are available through commercial suppliers.

25 To direct a Zsig9 polypeptide into the secretory pathway of a host cell, a secretory signal sequence (also known as a leader sequence, prepro sequence or pre sequence) is provided in the expression vector. The secretory signal sequence may be that of the protein, or
30 may be derived from another secreted protein (e.g., t-PA) or synthesized *de novo*. The secretory signal sequence is joined to the Zsig9 DNA sequence in the correct reading frame. Secretory signal sequences are commonly positioned 5' to the DNA sequence encoding the polypeptide of
35 interest, although certain signal sequences may be positioned elsewhere in the DNA sequence of interest (see,

e.g., Welch et al., U.S. Patent No. 5,037,743; Holland et al., U.S. Patent No. 5,143,830).

Cultured mammalian cells are preferred hosts within
5 the present invention. Methods for introducing exogenous DNA into mammalian host cells include calcium phosphate-mediated transfection, Wigler et al., *Cell* 14:725, (1978); Corsaro and Pearson, *Somatic Cell Genetics* 7:603, (1981); Graham and Van der Eb, *Virology* 52:456, (1973),
10 electroporation, Neumann et al., *EMBO J.* 1:841-845, (1982), DEAE-dextran mediated transfection, Ausubel et al., eds., *Current Protocols in Molecular Biology*, John Wiley and Sons, Inc., NY, (1987), and liposome-mediated transfection, Hawley-Nelson et al., *Focus* 15:73, (1993);
15 Ciccarone et al., *Focus* 15:80, (1993). The production of recombinant polypeptides in cultured mammalian cells is disclosed, for example, by Levinson et al., U.S. Patent No. 4,713,339; Hagen et al., U.S. Patent No. 4,784,950; Palmiter et al., U.S. Patent No. 4,579,821; and Ringold,
20 U.S. Patent No. 4,656,134. Suitable cultured mammalian cells include the COS-1 (ATCC No. CRL 1650), COS-7 (ATCC No. CRL 1651), BHK (ATCC No. CRL 1632), BHK 570 (ATCC No. CRL 10314), 293, ATCC No. CRL 1573; Graham et al., *J. Gen. Virol.* 36:59-72, (1977) and Chinese hamster ovary (e.g.
25 CHO-K1; ATCC No. CCL 61) cell lines. Additional suitable cell lines are known in the art and available from public depositories such as the American Type Culture Collection, Rockville, Maryland. In general, strong transcription promoters are preferred, such as promoters from SV-40 or
30 cytomegalovirus. See, e.g., U.S. Patent No. 4,956,288. Other suitable promoters include those from metallothionein genes (U.S. Patent Nos. 4,579,821 and 4,601,978, and the adenovirus major late promoter.

Drug selection is generally used to select for cultured mammalian cells into which foreign DNA has been inserted. Such cells are commonly referred to as "transfectants". Cells that have been cultured in the presence of the selective agent and are able to pass the gene of interest to their progeny are referred to as "stable transfectants." A preferred selectable marker is a gene encoding resistance to the antibiotic neomycin. Selection is carried out in the presence of a neomycin-type drug, such as G-418 or the like. Selection systems may also be used to increase the expression level of the gene of interest, a process referred to as "amplification." Amplification is carried out by culturing transfectants in the presence of a low level of the selective agent and then increasing the amount of selective agent to select for cells that produce high levels of the products of the introduced genes. A preferred amplifiable selectable marker is dihydrofolate reductase, which confers resistance to methotrexate. Other drug resistance genes (e.g. hygromycin resistance, multi-drug resistance, puromycin acetyltransferase) can also be used.

Other higher eukaryotic cells can also be used as hosts, including insect cells, plant cells and avian cells. Transformation of insect cells and production of foreign polypeptides therein is disclosed by Guarino et al., U.S. Patent No. 5,162,222; Bang et al., U.S. Patent No. 4,775,624; and WIPO publication WO 94/06463. The use of *Agrobacterium rhizogenes* as a vector for expressing genes in plant cells has been reviewed by Sinkar et al., *J. Biosci. (Bangalore)* 11:47-58, (1987).

Fungal cells, including yeast cells, and particularly cells of the genus *Saccharomyces*, can also be used within the present invention, such as for producing protein fragments or polypeptide fusions. Methods for transforming yeast cells with exogenous DNA and producing recombinant polypeptides therefrom are disclosed by, for example, Kawasaki, U.S. Patent No. 4,599,311; Kawasaki et al., U.S. Patent No. 4,931,373; Brake, U.S. Patent No. 4,870,008; Welch et al., U.S. Patent No. 5,037,743; and Murray et al., U.S. Patent No. 4,845,075. Transformed cells are selected by phenotype determined by the selectable marker, commonly drug resistance or the ability to grow in the absence of a particular nutrient (e.g., leucine). A preferred vector system for use in yeast is the *POT1* vector system disclosed by Kawasaki et al. (U.S. Patent No. 4,931,373), which allows transformed cells to be selected by growth in glucose-containing media. Suitable promoters and terminators for use in yeast include those from glycolytic enzyme genes (see, e.g., Kawasaki, U.S. Patent No. 4,599,311; Kingsman et al., U.S. Patent No. 4,615,974; and Bitter, U.S. Patent No. 4,977,092) and alcohol dehydrogenase genes. See also U.S. Patents Nos. 4,990,446; 5,063,154; 5,139,936 and 4,661,454. Transformation systems for other yeasts, including *Hansenula polymorpha*, *Schizosaccharomyces pombe*, *Kluyveromyces lactis*, *Kluyveromyces fragilis*, *Ustilago maydis*, *Pichia pastoris*, *Pichia methanolica*, *Pichia guilliermondii* and *Candida maltosa* are known in the art. See, for example, Gleeson et al., *J. Gen. Microbiol.* 132:3459-3465, (1986) and Cregg, U.S. Patent No. 4,882,279.

Aspergillus cells may be utilized according to the methods of McKnight et al., U.S. Patent No. 4,935,349.

Methods for transforming *Acremonium chrysogenum* are disclosed by Sumino et al., U.S. Patent No. 5,162,228.

Methods for transforming *Neurospora* are disclosed by Lambowitz, U.S. Patent No. 4,486,533.

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Transformed or transfected host cells are cultured according to conventional procedures in a culture medium containing nutrients and other components required for the growth of the chosen host cells. A variety of suitable media, including defined media and complex media, are known in the art and generally include a carbon source, a nitrogen source, essential amino acids, vitamins and minerals. Media may also contain such components as growth factors or serum, as required. The growth medium will generally select for cells containing the exogenously added DNA by, for example, drug selection or deficiency in an essential nutrient which is complemented by the selectable marker carried on the expression vector or co-transfected into the host cell.

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Within one aspect of the present invention, a novel protein is produced by a cultured cell, and the cell is used to screen for a receptor or receptors for the protein, including the natural receptor, as well as agonists and antagonists of the natural ligand.

Proteins of the present invention are useful for enhancing the growth or development of the placenta, heart, and liver. Zsig9 can be measured *in vitro* using cultured cells or *in vivo* by administering molecules of the claimed invention to the appropriate animal model. For instance, Zsig9 transfected (or co-transfected) expression host cells may be embedded in an alginate environment and injected (implanted) into recipient animals. Alginate-poly-L-lysine microencapsulation, permselective membrane encapsulation and diffusion

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chambers have been described as a means to entrap transfected mammalian cells or primary mammalian cells. These types of non-immunogenic "encapsulations" or microenvironments permit the transfer of nutrients into
5 the microenvironment, and also permit the diffusion of proteins and other macromolecules secreted or released by the captured cells across the environmental barrier to the recipient animal. Most importantly, the capsules or microenvironments mask and shield the foreign, embedded
10 cells from the recipient animal's immune response. Such microenvironments can extend the life of the injected cells from a few hours or days (naked cells) to several weeks (embedded cells).

15 Alginate threads provide a simple and quick means for generating embedded cells. The materials needed to generate the alginate threads are readily available and relatively inexpensive. Once made, the alginate threads are relatively strong and durable, both *in vitro* and,
20 based on data obtained using the threads, *in vivo*. The alginate threads are easily manipulable and the methodology is scaleable for preparation of numerous threads. In an exemplary procedure, 3% alginate is prepared in sterile H₂O, and sterile filtered. Just prior
25 to preparation of alginate threads, the alginate solution is again filtered. An approximately 50% cell suspension (containing about 5×10^5 to about 5×10^7 cells/ml) is mixed with the 3% alginate solution. One ml of the alginate/cell suspension is extruded into a 100 mM sterile
30 filtered CaCl₂ solution over a time period of ~15 min, forming a "thread". The extruded thread is then transferred into a solution of 50 mM CaCl₂, and then into a solution of 25 mM CaCl₂. The thread is then rinsed with deionized water before coating the thread by incubating in
35 a 0.01% solution of poly-L-lysine. Finally, the thread is rinsed with Lactated Ringer's Solution and drawn from

solution into a syringe barrel (without needle attached). A large bore needle is then attached to the syringe, and the thread is intraperitoneally injected into a recipient in a minimal volume of the Lactated Ringer's Solution.

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An alternative *in vivo* approach for assaying proteins of the present invention involves viral delivery systems. Exemplary viruses for this purpose include adenovirus, herpesvirus, vaccinia virus and adeno-associated virus (AAV). Adenovirus, a double-stranded DNA virus, is currently the best studied gene transfer vector for delivery of heterologous nucleic acid (for a review, see T.C. Becker et al., *Meth. Cell Biol.* 43:161-89 (1994); and J.T. Douglas and D.T. Curiel, *Science & Medicine* 4:44-53 (1997). The adenovirus system offers several advantages: adenovirus can (i) accommodate relatively large DNA inserts; (ii) be grown to high-titer; (iii) infect a broad range of mammalian cell types; and (iv) be used with a large number of available vectors containing different promoters. Also, because adenoviruses are stable in the bloodstream, they can be administered by intravenous injection. Some disadvantages (especially for gene therapy) associated with adenovirus gene delivery include: (i) very low efficiency integration into the host genome; (ii) existence in primarily episomal form; and (iii) the host immune response to the administered virus, precluding readministration of the adenoviral vector.

By deleting portions of the adenovirus genome, larger inserts (up to 7 kb) of heterologous DNA can be accommodated. These inserts may be incorporated into the viral DNA by direct ligation or by homologous recombination with a co-transfected plasmid. In an exemplary system, the essential E1 gene has been deleted from the viral vector, and the virus will not replicate unless the E1 gene is provided by the host cell (i.e., the

human 293 cell line). When intravenously administered to intact animals, adenovirus primarily targets the liver. If the adenoviral delivery system has an E1 gene deletion, the virus cannot replicate in the host cells. However, the host's tissue (i.e., liver) will express and process (and, if a signal sequence is present, secrete) the heterologous protein. Secreted proteins will enter the circulation in the highly vascularized liver, and effects on the infected animal can be determined.

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The adenovirus system can also be used for protein production *in vitro*. By culturing adenovirus-infected non-293 cells under conditions where the cells are not rapidly dividing, the cells can produce proteins for extended periods of time. For instance, BHK cells are grown to confluence in cell factories, then exposed to the adenoviral vector encoding the secreted protein of interest. The cells are then grown under serum-free conditions, which allows infected cells to survive for several weeks without significant cell division. Alternatively, adenovirus vector infected 293S cells can be grown in suspension culture at relatively high cell density to produce significant amounts of protein (see A. Garnier et al., *Cytotechnol.* 15:145-55 (1994)). With either protocol, an expressed, secreted heterologous protein can be repeatedly isolated from the cell culture supernatant. Within the infected 293S cell production protocol, non-secreted proteins may also be effectively obtained.

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PROTEIN ISOLATION:

Expressed recombinant polypeptides (or chimeric polypeptides) can be purified using fractionation and/or conventional purification methods and media. Ammonium sulfate precipitation and acid or chaotrope extraction may

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be used for fractionation of samples. Exemplary purification steps may include hydroxyapatite, size exclusion, FPLC and reverse-phase high performance liquid chromatography. Suitable anion exchange media include derivatized dextrans, agarose, cellulose, polyacrylamide, specialty silicas, and the like. PEI, DEAE, QAE and Q derivatives are preferred, with DEAE Fast-Flow Sepharose (Pharmacia, Piscataway, NJ) being particularly preferred. Exemplary chromatographic media include those media derivatized with phenyl, butyl, or octyl groups, such as Phenyl-Sepharose FF (Pharmacia), Toyopearl butyl 650 (Toso Haas, Montgomeryville, PA), Octyl-Sepharose (Pharmacia) and the like; or polyacrylic resins, such as Amberchrom CG 71 (Toso Haas) and the like. Suitable solid supports include glass beads, silica-based resins, cellulosic resins, agarose beads, cross-linked agarose beads, polystyrene beads, cross-linked polyacrylamide resins and the like that are insoluble under the conditions in which they are to be used. These supports may be modified with reactive groups that allow attachment of proteins by amino groups, carboxyl groups, sulfhydryl groups, hydroxyl groups and/or carbohydrate moieties. Examples of coupling chemistries include cyanogen bromide activation, N-hydroxysuccinimide activation, epoxide activation, sulfhydryl activation, hydrazide activation, and carboxyl and amino derivatives for carbodiimide coupling chemistries. These and other solid media are well known and widely used in the art, and are available from commercial suppliers. Methods for binding receptor polypeptides to support media are well known in the art. Selection of a particular method is a matter of routine design and is determined in part by the properties of the chosen support. See, for example, *Affinity Chromatography: Principles & Methods*, Pharmacia LKB Biotechnology, Uppsala, Sweden, (1988).

The polypeptides of the present invention can be isolated by exploitation of their properties. For example, immobilized metal ion adsorption (IMAC) chromatography can be used to purify histidine-rich proteins. Briefly, a gel is first charged with divalent metal ions to form a chelate, E. Sulkowski, *Trends in Biochem.* 3:1-7, (1985). Histidine-rich proteins will be adsorbed to this matrix with differing affinities, depending upon the metal ion used, and will be eluted by competitive elution, lowering the pH, or use of strong chelating agents. Other methods of purification include purification of glycosylated proteins by lectin affinity chromatography and ion exchange chromatography, *Methods in Enzymol.*, Vol. 182, "Guide to Protein Purification", M. Deutscher, (ed.), Acad. Press, San Diego, (1990), pp.529-39. Alternatively, a fusion of the polypeptide of interest and an affinity tag (e.g., polyhistidine, maltose-binding protein, an immunoglobulin domain) may be constructed to facilitate purification.

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Uses

Overexpression of Zsig9 in tumor cells indicates that Zsig9 can be used as an indicator for cancer. Thus, antibodies of Zsig9 can be used as a diagnostic to determine the presence of Zsig9 and thus the presence of cancer as explained below. Furthermore, antibodies labeled with radioisotopes or fused with toxins can be used as a therapeutic to treat cancer. Anti-sense nucleotides derived from the Zsig9 cDNA can also be used as a therapeutic to inhibit the growth of tumor cells.

Suitable detectable molecules may be directly or indirectly attached to the polypeptide or antibody, and include radionuclides, enzymes, substrates, cofactors,

inhibitors, fluorescent markers, chemiluminescent markers, magnetic particles and the like. Suitable cytotoxic molecules may be directly or indirectly attached to the polypeptide or antibody, and include bacterial or plant
5 toxins (for instance, diphtheria toxin, *Pseudomonas* exotoxin, ricin, abrin and the like), as well as therapeutic radionuclides, such as iodine-131, rhenium-188 or yttrium-90 (either directly attached to the polypeptide or antibody, or indirectly attached through means of a
10 chelating moiety, for instance). Polypeptides or antibodies may also be conjugated to cytotoxic drugs, such as adriamycin. For indirect attachment of a detectable or cytotoxic molecule, the detectable or cytotoxic molecule may be conjugated with a member of a complementary/
15 anticomplementary pair, where the other member is bound to the polypeptide or antibody portion. For these purposes, biotin/streptavidin is an exemplary complementary/ anticomplementary pair.

20 In another embodiment, polypeptide-toxin fusion proteins or antibody/fragment-toxin fusion proteins may be used for targeted cell or tissue inhibition or ablation (for instance, to treat cancer cells or tissues). Alternatively, if the polypeptide has multiple functional
25 domains (i.e., an activation domain or a ligand binding domain, plus a targeting domain), a fusion protein including only the targeting domain may be suitable for directing a detectable molecule, a cytotoxic molecule or a complementary molecule to a cell or tissue type of
30 interest. In instances where the domain only fusion protein includes a complementary molecule, the anticomplementary molecule may be conjugated to a detectable or cytotoxic molecule. Such domain-complementary molecule fusion proteins thus represent a generic targeting vehicle
35 for cell/tissue-specific delivery of generic anticomplementary-detectable/ cytotoxic molecule conjugates.

In another embodiment, polypeptide-cytokine fusion proteins or antibody/fragment-cytokine fusion proteins may be used for enhancing *in vitro* cytotoxicity (for instance, that mediated by monoclonal antibodies against tumor targets) and for enhancing *in vivo* killing of target tissues (for example, blood and bone marrow cancers). See, generally, J.L. Hornick et al., Blood 89:4437-47 (1997). In general, cytokines are toxic if administered systemically. The described fusion proteins enable targeting of a cytokine to a desired site of action, thereby providing an elevated local concentration of cytokine. Suitable Zsig9 polypeptides or anti-Zsig9 antibodies target an undesirable cell or tissue (i.e., a tumor or a leukemia), and the fused cytokine mediates improved target cell lysis by effector cells. Suitable cytokines for this purpose include interleukin 2 and granulocyte-macrophage colony-stimulating factor (GM-CSF), for instance.

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In yet another embodiment, if the Zsig9 polypeptide or anti-Zsig9 antibody targets vascular cells or tissues, such polypeptide or antibody may be conjugated with a radionuclide, and particularly with a beta-emitting radionuclide, to reduce restenosis. Such therapeutic approach poses less danger to clinicians who administer the radioactive therapy. For instance, iridium-192 impregnated ribbons placed into stented vessels of patients until the required radiation dose was delivered showed decreased tissue growth in the vessel and greater luminal diameter than the control group, which received placebo ribbons. Further, revascularisation and stent thrombosis were significantly lower in the treatment group. Similar results are predicted with targeting of a bioactive conjugate containing a radionuclide, as described herein.

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The bioactive polypeptide or antibody conjugates described herein can be delivered intravenously, intraarterially or intraductally, or may be introduced
5 locally at the intended site of action.

USES OF POLYNUCLEOTIDE/POLYPEPTIDE:

Molecules of the present invention can be used to
10 identify and isolate receptors of Zsig9. For example, proteins and peptides of the present invention can be immobilized on a column and membrane preparations run over the column (*Immobilized Affinity Ligand Techniques*, Hermanson et al., eds., pp.195-202 (Academic Press, San
15 Diego, CA, 1992). Proteins and peptides can also be radiolabeled (*Methods in Enzymol.*, vol. 182: "Guide to Protein Purification", M. Deutscher, ed., 721-737 (Acad. Press, San Diego, 1990,) or photoaffinity labeled (Brunner et al., *Ann. Rev. Biochem.* 62:483-514 (1993 and Fedan et
20 al., *Biochem. Pharmacol.* 33:1167-1180 (1984) and specific cell-surface proteins can be identified.

Another aspect of the present invention involves antisense polynucleotide compositions that are
25 complementary to a segment of the polynucleotides set forth in SEQ ID NOs: 1, 16 or 18. Such synthetic antisense oligonucleotides are designed to bind to mRNA encoding Zsig9 polypeptides and to inhibit translation of such mRNA. Such antisense oligonucleotides are used to
30 inhibit expression of Zsig9 polypeptide-encoding genes in cell culture or in a subject.

The present invention also provides reagents which will find use in diagnostic applications. For example,
35 the Zsig9 gene, a probe comprising Zsig9 DNA or RNA or a

subsequence thereof can be used to determine if the zZsig9 gene is present on chromosome 12 or if a mutation has occurred. Detectable chromosomal aberrations at the Zsig9 gene locus include, but are not limited to, aneuploidy, gene copy number changes, insertions, deletions, restriction site changes and rearrangements. Such aberrations can be detected using polynucleotides of the present invention by employing molecular genetic techniques, such as restriction fragment length polymorphism (RFLP) analysis, short tandem repeat (STR) analysis employing PCR techniques, and other genetic linkage analysis techniques known in the art (Sambrook et al., *ibid.*; Ausubel et. al., *ibid.*; A.J. Marian, Chest 108:255-65, 1995).

Transgenic mice, engineered to express the Zsig9 gene, and mice that exhibit a complete absence of Zsig9 gene function, referred to as "knockout mice", Snouwaert et al., *Science* 257:1083 (1992), may also be generated, Lowell et al., *Nature* 366:740-42 (1993). These mice may be employed to study the Zsig9 gene and the protein encoded thereby in an *in vivo* system.

CHROMOSOMAL LOCALIZATION:

Radiation hybrid mapping is a somatic cell genetic technique developed for constructing high-resolution, contiguous maps of mammalian chromosomes, Cox et al., *Science* 250:245-50 (1990). Partial or full knowledge of a gene's sequence allows one to design PCR primers suitable for use with chromosomal radiation hybrid mapping panels. Commercially available radiation hybrid mapping panels which cover the entire human genome, such as the Stanford G3 RH Panel and the GeneBridge 4 RH Panel (Research

Genetics, Inc., Huntsville, AL), are available. These panels enable rapid, PCR-based chromosomal localizations and ordering of genes, sequence-tagged sites (STSs), and other nonpolymorphic and polymorphic markers within a region of interest. This includes establishing directly proportional physical distances between newly discovered genes of interest and previously mapped markers. The precise knowledge of a gene's position can be useful for a number of purposes, including: 1) determining if a sequence is part of an existing contig and obtaining additional surrounding genetic sequences in various forms, such as YACs, BACs or cDNA clones; 2) providing a possible candidate gene for an inheritable disease which shows linkage to the same chromosomal region; and 3) cross-referencing model organisms, such as mouse, which may aid in determining what function a particular gene might have.

For pharmaceutical use, the proteins of the present invention are formulated for parenteral, particularly intravenous or subcutaneous, delivery according to conventional methods. Intravenous administration will be by bolus injection or infusion over a typical period of one to several hours. In general, pharmaceutical formulations will include a Zsig9 protein in combination with a pharmaceutically acceptable vehicle, such as saline, buffered saline, 5% dextrose in water or the like. Formulations may further include one or more excipients, preservatives, solubilizers, buffering agents, albumin to prevent protein loss on vial surfaces, etc. Methods of formulation are well known in the art and are disclosed, for example, in *Remington's Pharmaceutical Sciences*, 19th Edition Gennaro, ed., (Mack Publishing Co., Easton PA, 1995). Therapeutic doses will generally be in the range of 0.1 to 100 lg/kg of patient weight per day, preferably 0.5-20 lg/kg per day, with the exact dose determined by

the clinician according to accepted standards, taking into account the nature and severity of the condition to be treated, patient traits, etc. Determination of dose is within the level of ordinary skill in the art. The proteins may be administered for acute treatment, over one week or less, often over a period of one to three days or may be used in chronic treatment, over several months or years. In general, a therapeutically effective amount of Zsig9 is an amount sufficient to produce a clinically significant change in the liver, kidney, heart or placenta.

Zsig9 Structure

The potential N-terminal 3 dimensional structure of Zsig9 should have similarities to that predicted for amylin and calcitonin-gene related peptides (CGRP). Moving from the N-terminus to the C-terminus there is a predicted disulfide bond between residues 28 and 31 of SEQ ID NO:2. This is like the predicted disulfide of amylin (positions 35-40) and CGRP1 and 2 (positions 84-89). If the mature form of Zsig9 is not cleaved at the Lys-Lys at position 48-49, there is probably a beta turn followed by an extended region with hydrophobic residues packing along the hydrophobic face of the helix. The C-terminus is probably not amidated as in amylin and CGRP if the mature form is cleaved at the Lys-Lys at position 48-49. If the mature form ends at either the serine at position 81 or at the phenylalanine at position 83, then the glycine at position 82 or the glycine at position 84 may be used to amidate the C-terminus.

Zsig9 Tissue Distribution/Multiple mRNA sizes

Northern analysis shows that Zsig9 is ubiquitous with higher levels in placenta, pancreas, thyroid, prostate, and liver. This may indicate that it is involved in homeostasis. Zsig9 is transcribed by many cell types and is probably released by a variety of cell types in response to some metabolic stress. The Zsig9 receptor is then activated by Zsig9 to alleviate the stress. The stress may include non-optimal levels of sugar, carbohydrate, antigenic load, fat, temperature, pH, O₂, osmotic concentration and the like.

Zsig9 is overexpressed in tumor cells. Thus, antibodies to Zsig9 can be used to diagnosis the presence of tumors. Radiolabeled antibodies to Zsig9 can further be used as an imaging agent to locate and treat tumors in the body. Anti-sense nucleotides to *zsig9* can be used to inhibit the progression of tumors.

Therapeutic utilityCancer Diagnosis and Therapy

As can be seen in Example 5, Zsig9 is overexpressed in a number of human tumors including brain, liver, lung, esophageal, stomach, colon, rectal, thyroid, and lymphoma tumors. Thus, antibodies to Zsig9 can be used both to detect and treat the tumors which overexpress Zsig9. Radiolabeled antibodies to Zsig9 can used both to detect and localize tumors. Antibodies which are either radiolabeled or to which a toxic polypeptide is fused can also be used to treat tumors which overexpress Zsig9.

Nucleotide primers and probes of the *Zsig9* gene can be used to detect the overexpression of *Zsig9* using PCR. Gene amplification of *Zsig9* can be determined indirectly by assay of a patient body fluid to detect the presence of elevated levels of *Zsig9*. Suitable body fluids include serum and urine and exudates of the putative tumor tissues. Examples of immunoassays which can be used in determining the expression of *Zsig9* to diagnose neoplastic diseases are described in the patent and scientific literature. See, for example U.S. Patent Nos. 3,791,932; 3,817,837; 3,839,153; 3,850,752; 3,850,578; 3,853,987; 3,867,517; 3,879,262; 3,901,654; 3,935,074; 3,984,533; 3,996,345; 4,034,074; and 4,098,876. Antisense nucleotides to the *Zsig9* DNA and RNA can be administered to a patient to inhibit expression of *Zsig9* and thus inhibit tumor growth. Methods for

Antibodies to the *Zsig9* polypeptide can be purified and then administered to a patient. These reagents can be combined for therapeutic use with additional active or inert ingredients, e.g., in pharmaceutically acceptable carriers or diluents along with physiologically innocuous stabilizers and excipients. These combinations can be sterile filtered and placed into dosage forms as by lyophilization in dosage vials or storage in stabilized aqueous preparations. This invention also contemplates use of antibodies, binding fragments thereof or single-chain antibodies of the antibodies including forms which are not complement binding.

30

The quantities of reagents necessary for effective therapy will depend upon many different factors, including means of administration, target site, physiological state of the patient, and other medications administered. Thus, treatment dosages should be titrated to optimize safety

and efficacy. Typically, dosages used *in vitro* may provide useful guidance in the amounts useful for *in vivo* administration of these reagents. Animal testing of effective doses for treatment of particular disorders will provide further predictive indication of human dosage. Methods for administration include oral, intravenous, peritoneal, intramuscular, or transdermal administration. Pharmaceutically acceptable carriers will include water, saline, buffers to name just a few. Dosage ranges would ordinarily be expected from 1 μ g to 1000 μ g per kilogram of body weight per day. However, the doses may be higher or lower as can be determined by a medical doctor with ordinary skill in the art. For a complete discussion of drug formulations and dosage ranges see *Remington's Pharmaceutical Sciences*, 19th Ed., (Mack Publishing Co., Easton, Penn., 1995), and *Goodman and Gilman's: The Pharmacological Bases of Therapeutics*, 9th Ed. (Pergamon Press 1996).

20 Nucleic Acid-based Therapeutic Treatment

If a mammal has a mutated or lacks a Zsig9 gene, the Zsig9 gene can be introduced into the cells of the mammal. In one embodiment, a gene encoding a Zsig9 polypeptide is introduced *in vivo* in a viral vector. Such vectors include an attenuated or defective DNA virus, such as but not limited to herpes simplex virus (HSV), papillomavirus, Epstein Barr virus (EBV), adenovirus, adeno-associated virus (AAV), and the like. Defective viruses, which entirely or almost entirely lack viral genes, are preferred. A defective virus is not infective after introduction into a cell. Use of defective viral vectors allows for administration to cells in a specific, localized area, without concern that the vector can infect other cells. Examples of particular vectors include, but

are not limited to, a defective herpes virus 1 (HSV1) vector [Kaplitt et al., *Molec. Cell. Neurosci.*, 2 :320-330 (1991)], an attenuated adenovirus vector, such as the vector described by Stratford-Perricaudet et al., *J. Clin. Invest.*, 90 :626-630 (1992), and a defective adeno-associated virus vector [Samulski et al., *J. Virol.*, 61:3096-3101 (1987); Samulski et al. *J. Virol.*, 63:3822-3828 (1989)].

10 In another embodiment, the gene can be introduced in a retroviral vector, e.g., as described in Anderson et al., U.S. Patent No. 5,399,346; Mann et al., *Cell*, 33:153 (1983); Temin et al., U.S. Patent No. 4,650,764; Temin et al., U.S. Patent No. 4,980,289; Markowitz et al., *J. Virol.*, 62:1120 (1988); Temin et al., U.S. Patent No. 5,124,263; International Patent Publication No. WO 95/07358, published March 16, 1995 by Dougherty et al.; and Blood, 82:845 (1993).

20 Alternatively, the vector can be introduced by lipofection *in vivo* using liposomes. Synthetic cationic lipids can be used to prepare liposomes for *in vivo* transfection of a gene encoding a marker [Felgner et al., *Proc. Natl. Acad. Sci. USA*, 84:7413-7417 (1987); see
25 Mackey et al., *Proc. Natl. Acad. Sci. USA*, 85:8027-8031 (1988)]. The use of lipofection to introduce exogenous genes into specific organs *in vivo* has certain practical advantages. Molecular targeting of liposomes to specific cells represents one area of benefit. It is clear that
30 directing transfection to particular cells represents one area of benefit. It is clear that directing transfection to particular cell types would be particularly advantageous in a tissue with cellular heterogeneity, such

as the pancreas, liver, kidney, and brain. Lipids may be chemically coupled to other molecules for the purpose of targeting. Targeted peptides, e.g., hormones or neurotransmitters, and proteins such as antibodies, or
5 non-peptide molecules could be coupled to liposomes chemically.

It is possible to remove the cells from the body and introduce the vector as a naked DNA plasmid and then re-
10 implant the transformed cells into the body. Naked DNA vector for gene therapy can be introduced into the desired host cells by methods known in the art, e.g., transfection, electroporation, microinjection, transduction, cell fusion, DEAE dextran, calcium phosphate
15 precipitation, use of a gene gun or use of a DNA vector transporter [see, e.g., Wu et al., *J. Biol. Chem.*, 267:963-967 (1992); Wu et al., *J. Biol. Chem.*, 263:14621-14624 (1988)].

20 Zsig9 polypeptides can also be used to prepare antibodies that specifically bind to Zsig9 epitopes, peptides or polypeptides. The Zsig9 polypeptide or a fragment thereof is inoculate into an animal so as to elicit an immune response. Antibodies generated from this
25 immune response can be isolated and purified as described herein. Methods for preparing and isolating polyclonal and monoclonal antibodies are well known in the art. See, for example, *Current Protocols in Immunology*, Cooligan, et al., Eds., (National Institutes of Health, John Wiley and
30 Sons, Inc., 1995); Sambrook et al., *Molecular Cloning: A Laboratory Manual, Second Edition*, (Cold Spring Harbor, NY, 1989); and Hurrell, J. G. R., Ed., *Monoclonal Hybridoma Antibodies: Techniques and Applications*, (CRC Press, Inc., Boca Raton, FL, 1982).

As would be evident to one of ordinary skill in the art, polyclonal antibodies can be generated from inoculating a variety of warm-blooded animals such as horses, cows, goats, sheep, dogs, chickens, rabbits, mice, and rats with a Zsig9 polypeptide or a fragment thereof. The immunogenicity of a Zsig9 polypeptide may be increased through the use of an adjuvant, such as alum (aluminum hydroxide) or Freund's complete or incomplete adjuvant. Polypeptides useful for immunization also include fusion polypeptides, such as fusions of Zsig9 or a portion thereof with an immunoglobulin polypeptide or with maltose binding protein. The polypeptide immunogen may be a full-length molecule or a portion thereof. If the polypeptide portion is "hapten-like", such portion may be advantageously joined or linked to a macromolecular carrier (such as keyhole limpet hemocyanin (KLH), bovine serum albumin (BSA) or tetanus toxoid) for immunization.

As used herein, the term "antibodies" includes polyclonal antibodies, affinity-purified polyclonal antibodies, monoclonal antibodies, and antigen-binding fragments, such as F(ab')₂ and Fab proteolytic fragments. Genetically engineered intact antibodies or fragments, such as chimeric antibodies, Fv fragments, single chain antibodies and the like, as well as synthetic antigen-binding peptides and polypeptides, are also included. Non-human antibodies may be humanized by grafting non-human CDRs onto human framework and constant regions, or by incorporating the entire non-human variable domains (optionally "cloaking" them with a human-like surface by replacement of exposed residues, wherein the result is a "veneered" antibody). In some instances, humanized antibodies may retain non-human residues within the human

variable region framework domains to enhance proper binding characteristics. Through humanizing antibodies, biological half-life may be increased, and the potential for adverse immune reactions upon administration to humans is reduced.

Alternative techniques for generating or selecting antibodies useful herein include *in vitro* exposure of lymphocytes to Zsig9 protein or peptide, and selection of antibody display libraries in phage or similar vectors (for instance, through use of immobilized or labeled Zsig9 protein or peptide). Genes encoding polypeptides having potential Zsig9 polypeptide binding domains can be obtained by screening random peptide libraries displayed on phage (phage display) or on bacteria, such as *E. coli*. Nucleotide sequences encoding the polypeptides can be obtained in a number of ways, such as through random mutagenesis and random polynucleotide synthesis. These random peptide display libraries can be used to screen for peptides which interact with a known target which can be a protein or polypeptide, such as a ligand or receptor, a biological or synthetic macromolecule, or organic or inorganic substances.

Techniques for creating and screening such random peptide display libraries are known in the art (Ladner et al., US Patent NO. 5,223,409; Ladner et al., US Patent NO. 4,946,778; Ladner et al., US Patent NO. 5,403,484 and Ladner et al., US Patent NO. 5,571,698) and random peptide display libraries and kits for screening such libraries are available commercially, for instance from Clontech (Palo Alto, CA), Invitrogen Inc. (San Diego, CA), New England Biolabs, Inc. (Beverly, MA) and Pharmacia LKB Biotechnology Inc. (Piscataway, NJ). Random peptide

display libraries can be screened using the Zsig9 sequences disclosed herein to identify proteins which bind to Zsig9. These "binding proteins" which interact with Zsig9 polypeptides can be used for tagging cells; for isolating homolog polypeptides by affinity purification; they can be directly or indirectly conjugated to drugs, toxins, radionuclides and the like. These binding proteins can also be used in analytical methods such as for screening expression libraries and neutralizing activity. The binding proteins can also be used for diagnostic assays for determining circulating levels of polypeptides; for detecting or quantitating soluble polypeptides as marker of underlying pathology or disease. These binding proteins can also act as Zsig9 "antagonists" to block Zsig9 binding and signal transduction *in vitro* and *in vivo*. These anti-Zsig9 binding proteins would be useful for inhibiting the growth of tumors.

Antibodies are determined to be specifically binding if: 1) they exhibit a threshold level of binding activity, and/or 2) they do not significantly cross-react with related polypeptide molecules. First, antibodies herein specifically bind if they bind to a Zsig9 polypeptide, peptide or epitope with a binding affinity (K_a) of 10^6 M^{-1} or greater, preferably 10^7 M^{-1} or greater, more preferably 10^8 M^{-1} or greater, and most preferably 10^9 M^{-1} or greater. The binding affinity of an antibody can be readily determined by one of ordinary skill in the art, for example, by Scatchard analysis, Scatchard, G., *Ann. NY Acad. Sci.* 51: 660-672 (1949).

Second, antibodies are determined to specifically bind if they do not significantly cross-react with related polypeptides. Antibodies do not significantly cross-react

with related polypeptide molecules, for example, if they detect Zsig9 but not known related polypeptides using a standard Western blot analysis (Ausubel et al., *ibid.*). Examples of known related polypeptides are orthologs, proteins from the same species that are members of a protein family (e.g. IL-16), Zsig9 polypeptides, and non-human Zsig9. Moreover, antibodies may be "screened against" known related polypeptides to isolate a population that specifically binds to the inventive polypeptides. For example, antibodies raised to Zsig9 are adsorbed to related polypeptides adhered to insoluble matrix; antibodies specific to Zsig9 will flow through the matrix under the proper buffer conditions. Such screening allows isolation of polyclonal and monoclonal antibodies non-crossreactive to closely related polypeptides, *Antibodies: A Laboratory Manual*, Harlow and Lane (eds.), (Cold Spring Harbor Laboratory Press, 1988); *Current Protocols in Immunology*, Cooligan, et al. (eds.), (National Institutes of Health, John Wiley and Sons, Inc., 1995). Screening and isolation of specific antibodies is well known in the art. See, *Fundamental Immunology*, Paul Eds., (Raven Press, 1993); Getzoff et al., *Adv. in Immunol.* 43: 1-98 (1988); *Monoclonal Antibodies: Principles and Practice*, Goding, J.W., Eds., (Academic Press Ltd., 1996); Benjamin et al., *Ann. Rev. Immunol.* 2: 67-101 (1984).

A variety of assays known to those skilled in the art can be utilized to detect antibodies which specifically bind to Zsig9 proteins or peptides. Exemplary assays are described in detail in *Antibodies: A Laboratory Manual*, Harlow and Lane, Eds., (Cold Spring Harbor Laboratory Press, 1988). Representative examples of such assays include: concurrent immunoelectrophoresis,

radioimmunoassay, radioimmuno-precipitation, enzyme-linked immunosorbent assay (ELISA), dot blot or Western blot assay, inhibition or competition assay, and sandwich assay. In addition, antibodies can be screened for
5 binding to wild-type versus mutant Zsig9 protein or polypeptide.

Antibodies to Zsig9 may be used for tagging cells that express Zsig9; for isolating Zsig9 by affinity
10 purification; for diagnostic assays for determining circulating levels of Zsig9 polypeptides; for detecting or quantitating soluble Zsig9 as marker of underlying pathology or disease; in analytical methods employing FACS; for screening expression libraries; for generating
15 anti-idiotypic antibodies; and as neutralizing antibodies or as antagonists to block Zsig9 *in vitro* and *in vivo*. Suitable direct tags or labels include radionuclides, enzymes, substrates, cofactors, inhibitors, fluorescent markers, chemiluminescent markers, magnetic particles and
20 the like; indirect tags or labels may feature use of biotin-avidin or other complement/anti-complement pairs as intermediates. Antibodies herein may also be directly or indirectly conjugated to drugs, toxins, radionuclides and the like, and these conjugates used for *in vivo* diagnostic
25 or therapeutic applications. Moreover, antibodies to Zsig9 or fragments thereof may be used *in vitro* to detect denatured Zsig9 or fragments thereof in assays, for example, Western Blots or other assays known in the art.

30 Another embodiment of the present invention provides for a peptide or polypeptide comprising an epitope-bearing portion of a polypeptide of the invention. The epitope of the this polypeptide portion is an immunogenic or antigenic epitope of a polypeptide of the invention. A

region of a protein to which an antibody can bind is defined as an "antigenic epitope". See for instance, Geysen, H.M. et al., *Proc. Natl. Acad Sci. USA* 81:3998-4002 (1984).

5

As to the selection of peptides or polypeptides bearing an antigenic epitope (i.e., that contain a region of a protein molecule to which an antibody can bind), it is well known in the art that relatively short synthetic
10 peptides that mimic part of a protein sequence are routinely capable of eliciting an antiserum that reacts with the partially mimicked protein. See Sutcliffe, J.G. et al. *Science* 219:660-666 (1983). Peptides capable of eliciting protein-reactive sera are frequently represented
15 in the primary sequence of a protein, can be characterized by a set of simple chemical rules, and are confined neither to immunodominant regions of intact proteins (i.e., immunogenic epitopes) nor to the amino or carboxyl terminals. Peptides that are extremely hydrophobic and
20 those of six or fewer residues generally are ineffective at inducing antibodies that bind to the mimicked protein; longer soluble peptides, especially those containing proline residues, usually are effective.

25 Antigenic epitope-bearing peptides and polypeptides of the invention are therefore useful to raise antibodies, including monoclonal antibodies, that bind specifically to a polypeptide of the invention. Antigenic epitope-bearing peptides and polypeptides of the
30 present invention contain a sequence of at least nine, preferably between 15 to about 30 amino acids contained within the amino acid sequence of a polypeptide of the invention. However, peptides or polypeptides comprising a larger portion of an amino acid sequence of the invention,
35 containing from 30 to 50 amino acids, or any length up to

and including the entire amino acid sequence of a polypeptide of the invention, also are useful for inducing antibodies that react with the protein. Preferably, the amino acid sequence of the epitope-bearing peptide is selected to provide substantial solubility in aqueous solvents (i.e., the sequence includes relatively hydrophilic residues and hydrophobic residues are preferably avoided); and sequences containing proline residues are particularly preferred. All of the polypeptides shown in the sequence listing contain antigenic epitopes to be used according to the present invention, however, specifically designed antigenic epitopes include the peptides defined by SEQ ID NO:24

15

BIOACTIVE CONJUGATES:

Antibodies or polypeptides herein can also be directly or indirectly conjugated to drugs, toxins, radionuclides and the like, and these conjugates used for *in vivo* diagnostic or therapeutic applications. For instance, polypeptides or antibodies of the present invention can be used to identify or treat tissues or organs that express a corresponding anti-complementary molecule (receptor or antigen, respectively, for instance). More specifically, Zsig9 polypeptides or anti-Zsig9 antibodies, or bioactive fragments or portions thereof, can be coupled to detectable or cytotoxic molecules and delivered to a mammal having cells, tissues or organs that express the anti-complementary molecule.

Suitable detectable molecules may be directly or indirectly attached to the polypeptide or antibody, and include radionuclides, enzymes, substrates, cofactors, inhibitors, fluorescent markers, chemiluminescent markers, magnetic particles and the like. Suitable cytotoxic

molecules may be directly or indirectly attached to the polypeptide or antibody, and include bacterial or plant toxins (for instance, diphtheria toxin, *Pseudomonas* exotoxin, ricin, abrin and the like), as well as
5 therapeutic radionuclides, such as iodine-131, rhenium-188 or yttrium-90 (either directly attached to the polypeptide or antibody, or indirectly attached through means of a chelating moiety, for instance). Polypeptides or
10 antibodies may also be conjugated to cytotoxic drugs, such as adriamycin. For indirect attachment of a detectable or cytotoxic molecule, the detectable or cytotoxic molecule can be conjugated with a member of a complementary/
15 anticomplementary pair, where the other member is bound to the polypeptide or antibody portion. For these purposes, biotin/streptavidin is an exemplary complementary/
anticomplementary pair.

In another embodiment, polypeptide-toxin fusion proteins or antibody-toxin fusion proteins can be used for
20 targeted cell or tissue inhibition or ablation (for instance, to treat cancer cells or tissues). Alternatively, if the polypeptide has multiple functional domains (i.e., an activation domain or a ligand binding domain, plus a targeting domain), a fusion protein
25 including only the targeting domain may be suitable for directing a detectable molecule, a cytotoxic molecule or a complementary molecule to a cell or tissue type of interest. In instances where the domain only fusion
protein includes a complementary molecule, the anti-
30 complementary molecule can be conjugated to a detectable or cytotoxic molecule. Such domain-complementary molecule fusion proteins thus represent a generic targeting vehicle for cell/tissue-specific delivery of generic anti-
complementary-detectable/ cytotoxic molecule conjugates.

In another embodiment, Zsig9-cytokine fusion proteins or antibody-cytokine fusion proteins can be used for enhancing *in vivo* killing of target tissues (for example, blood and bone marrow cancers), if the Zsig9 polypeptide or anti-Zsig9 antibody targets the hyperproliferative blood or bone marrow cell (See, generally, Hornick et al., *Blood* 89:4437-47 (1997)). They described fusion proteins enable targeting of a cytokine to a desired site of action, thereby providing an elevated local concentration of cytokine. Suitable Zsig9 polypeptides or anti-Zsig9 antibodies target an undesirable cell or tissue (i.e., a tumor or a leukemia), and the fused cytokine mediated improved target cell lysis by effector cells. Suitable cytokines for this purpose include interleukin-2 and granulocyte-macrophage colony-stimulating factor (GM-CSF), for instance.

In yet another embodiment, if the Zsig9 polypeptide or anti-Zsig9 antibody targets vascular cells or tissues, such polypeptide or antibody may be conjugated with a radionuclide, and particularly with a beta-emitting radionuclide, to reduce restenosis or to reduce the growth of blood vessels in tumors. Such therapeutic approach poses less danger to clinicians who administer the radioactive therapy.

The bioactive polypeptide or antibody conjugates described herein can be delivered intravenously, intraarterially or intraductally, or may be introduced locally at the intended site of action.

USES OF POLYNUCLEOTIDE/POLYPEPTIDE:

Molecules of the present invention can be used to identify and isolate receptors involved in the binding of Zsig9. For example, proteins and peptides of the

present invention can be immobilized on a column and membrane preparations run over the column, *Immobilized Affinity Ligand Techniques*, Hermanson et al., eds., pp.195-202 (Academic Press, San Diego, CA, 1992).

5 Proteins and peptides can also be radiolabeled *Methods in Enzymol.*, vol. 182, "Guide to Protein Purification", M. Deutscher, ed., p.p. 721-737 (Acad. Press, San Diego, 1990) or photoaffinity labeled, Brunner et al., *Ann. Rev. Biochem.* 62:483-514 1993 and Fedan et al., *Biochem.*

10 *Pharmacol.* 33:1167-80 (1984) and specific cell-surface proteins can be identified.

GENE THERAPY:

15 Polynucleotides encoding Zsig9 polypeptides are useful within gene therapy applications where it is desired to increase or inhibit Zsig9 activity. If a mammal has a mutated or absent Zsig9 gene, the Zsig9 gene can be introduced into the cells of the mammal. In one

20 embodiment, a gene encoding a Zsig9 polypeptide is introduced *in vivo* in a viral vector. Such vectors include an attenuated or defective DNA virus, such as, but not limited to, herpes simplex virus (HSV), papillomavirus, Epstein Barr virus (EBV), adenovirus,

25 adeno-associated virus (AAV), and the like. Defective viruses, which entirely or almost entirely lack viral genes, are preferred. A defective virus is not infective after introduction into a cell. Use of defective viral vectors allows for administration to cells in a specific,

30 localized area, without concern that the vector can infect other cells. Examples of particular vectors include, but are not limited to, a defective herpes simplex virus 1 (HSV1) vector (Kaplitt et al., *Molec. Cell. Neurosci.* 2:320-30 (1991); an attenuated adenovirus vector, such as

the vector described by Stratford-Perricaudet et al., *J. Clin. Invest.* 90:626-30 (1992); and a defective adeno-associated virus vector, Samulski et al., *J. Virol.* 61:3096-101 (1987); Samulski et al., *J. Virol.* 63:3822-8 (1989).

In another embodiment, a Zsig9 gene can be introduced in a retroviral vector, e.g., as described in Anderson et al., U.S. Patent No. 5,399,346; Mann et al. *Cell* 33:153 (1983); Temin et al., U.S. Patent No. 4,650,764; Temin et al., U.S. Patent No. 4,980,289; Markowitz et al., *J. Virol.* 62:1120 (1988); Temin et al., U.S. Patent No. 5,124,263; International Patent Publication No. WO 95/07358, published March 16, 1995 by Dougherty et al.; and Kuo et al., *Blood* 82:845 (1993).

Alternatively, the vector can be introduced by lipofection *in vivo* using liposomes. Synthetic cationic lipids can be used to prepare liposomes for *in vivo* transfection of a gene encoding a marker (Felgner et al., *Proc. Natl. Acad. Sci. USA* 84:7413-7 (1987); Mackey et al., *Proc. Natl. Acad. Sci. USA* 85:8027-31 (1988). The use of lipofection to introduce exogenous genes into specific organs *in vivo* has certain practical advantages. Molecular targeting of liposomes to specific cells represents one area of benefit. More particularly, directing transfection to particular cells represents one area of benefit. For instance, directing transfection to particular cell types would be particularly advantageous in a tissue with cellular heterogeneity, such as the pancreas, liver, kidney, and brain. Lipids may be chemically coupled to other molecules for the purpose of targeting. Targeted peptides (e.g., hormones or neurotransmitters), proteins such as antibodies, or non-peptide molecules can be coupled to liposomes chemically.

It is possible to remove the target cells from the body; to introduce the vector as a naked DNA plasmid; and then to re-implant the transformed cells into the body. Naked DNA vectors for gene therapy can be introduced into the desired host cells by methods known in the art, e.g., transfection, electroporation, microinjection, transduction, cell fusion, DEAE dextran, calcium phosphate precipitation, use of a gene gun or use of a DNA vector transporter. See, e.g., Wu et al., *J. Biol. Chem.* 267:963-7 (1992); Wu et al., *J. Biol. Chem.* 263:14621-4 (1988).

Antisense methodology can be used to inhibit Zsig9 gene transcription, such as to inhibit cell proliferation in vivo. Polynucleotides that are complementary to a segment of a Zsig9-encoding polynucleotide (e.g., a polynucleotide as set forth in SEQ ID NO:1) are designed to bind to Zsig9-encoding mRNA and to inhibit translation of such mRNA. Such antisense polynucleotides are used to inhibit expression of Zsig9 polypeptide-encoding genes in cell culture or in a subject.

The present invention also provides reagents which will find use in diagnostic applications. For example, the Zsig9 gene, a probe comprising Zsig9 DNA or RNA or a subsequence thereof can be used to determine if the Zsig9 gene is present on chromosome 12 or if a mutation has occurred. Detectable chromosomal aberrations at the Zsig9 gene locus include, but are not limited to, aneuploidy, gene copy number changes, insertions, deletions, restriction site changes and rearrangements. Such aberrations can be detected using polynucleotides of the present invention by employing molecular genetic

techniques, such as restriction fragment length polymorphism (RFLP) analysis, short tandem repeat (STR) analysis employing PCR techniques, and other genetic linkage analysis techniques known in the art (Sambrook et al., *ibid.*; Ausubel et. al., *ibid.*; Marian, *Chest* 108:255-65 (1995).

The invention is further illustrated by the following non-limiting examples.

Example 1. Cloning of Zsig9

Zsig9 was identified from expressed sequence tag (EST) SEQ ID NO: 7. The cDNA clone containing the EST was
5 discovered in a placenta from a full-term pregnancy cDNA library which contained the EST. The cDNA was isolated from *E. coli* transfected with the plasmid and then streaked out on an LB 100 µg/ml ampicillin and 100 µg/ml methicillin plate. The cDNA insert was sequenced. The
10 insert was determined to be 649 base pairs long with a 84 amino acid open reading frame and a putative 20 amino acid signal peptide.

Example 2

15

Construction of Zsig9 Expression Vectors

Two Zsig9 construction vectors were made in a flag amino acid sequence (SEQ ID NO: 8) was inserted onto the
20 N-terminal or C-terminal ends of the Zsig9 polypeptide. For the construction in which the flag amino acid sequence was attached to the N-terminus of Zsig9, a 473 bp Zsig9 PCR DNA fragment was generated with 1 µl of a ¼ dilution of the plasmid prep of Example 1 and 1 microliter (µl) of
25 SEQ ID NO: 9 and 10 each having a concentration of 20 picomoles (pm)/µl of primer. The PCR mixture contained 2.5 µl of 10X PCR buffer, 0.5 KLENTAQ (both from CLONTECH), 2.5 µl REDI-LOAD dye (Research Genetics), 2.5 nucleotide triphosphate mix (Perkin-Elmer) and 14 µl of water. The
30 PCR reaction was incubated at 94°C for 5 minutes, and then run for 10 cycles each individual cycle being comprised of

30 seconds at 94°C and 2 minutes at 75°C. This was followed by 15 cycles each cycle being comprised of 30 seconds at 94°C and 2 minutes at 60°C. The reaction was ended with an incubation for 10 minutes at 74°C.

5

The resultant PCR mixture was then run on a 0.9% LMP agarose gel with TBE buffer. After the gel was run the band containing the DNA was cut out and the DNA was purified from the gel with a QUIAQUICK® column (Qiagen).

10 100 µl of the DNA was digested in a solution containing 4 µl of B buffer, 1 µl of *Bam*HI (Boehringer Mannheim) and 1 µl of *Xho*I (Boehringer Mannheim) for 2 hours at 37°C. The digested reaction mixture was electrophoresed on a 1% TBE gel; the DNA band was excised with a razor blade and the
15 DNA was extracted from the gel with the Qiaquick® Gel Extraction Kit (Qiagen).

The excised DNA was subcloned into plasmid NF/pZP9 which had been cut with *Bam* and *Xho*. NF/pZP9 is a
20 mammalian cell expression vector comprising an expression cassette containing the mouse metallothionein-1 promoter, a sequence encoding the tissue plasminogen activator (TPA) leader, then the flag peptide (SEQ ID NO:8), then multiple restriction sites. These were followed by the human growth
25 hormone terminator, an *E. coli* origin of replication and a mammalian selectable marker expression unit containing the SV40 promoter, enhancer and origin of replication; a dihydrofolate reductase gene (DHFR) and the SV40 terminator. 1 µl containing 10 ng of the NF/pZP9 vector
30 which had been previously digested with *Xho* and *Bam*HI was mixed with 1 µl of 10X ligase buffer, 1 µl of T4 ligase and 2 µl of Zsig9 fragment containing 20 ng. The ligation took place at room temperature for 3 hours and then

electroporated into DH10b cells. After the electroporation the cells were plated onto LB-amp plates.

For the construction of the Zsig9 gene in which the
5 flag polypeptide SEQ ID NO: 8 was inserted onto the C-terminus of the Zsig9 polypeptide, a 649 bp Zsig9 PCR fragment was generated with 1 µl of ¼ dilution of the plasmid preparation containing Zsig9 described in Example 1 and 20 pm each of primers SEQ ID NO: 11 and SEQ ID NO:
10 12. The PCR reaction was incubated at 94°C for 5 minutes, then run for 10 cycles, each cycle being comprised 30 seconds at 94°C and 2 minutes at 75°C. This was followed by 15 cycles each cycle comprised of 30 seconds at 94°C and 2 minutes at 60°C. The reaction was ended with a final 10
15 minute extension at 74°C.

The entire reaction mixture was run on a 1% TBE gel and the DNA was cut out with a razor blade and the DNA was extracted using the QIAQUICK™ gel extraction kit. 20 µl
20 out of the recovered 35 µl digested with 10 units of *Bam*H1 (Boehringer Mannheim) and 10 units of *Eco*R1 (Gibco BRL) for 2 hours at 37°C. The digested PCR mixture was electrophoresed on a 1% TBE gel. The DNA band was cut out with a razor blade and the DNA was extracted from the gel
25 using the QIAquick® Gel Extraction Kit (Qiagen). The extracted DNA was subcloned into plasmid CF/pZP9 which had been cut with *Eco*R1 and *Bam*H1. Plasmid cfpzp9 is a mammalian expression vector containing an expression cassette having the mouse metallothionein-1 promoter,
30 multiple restriction sites for insertion of coding sequences, a sequence encoding the flag peptide, SEQ ID

NO:10, a stop codon, a human growth hormone terminator, an
E. coli origin of replication, a mammalian selectable
marker expression unit having an SV40 promoter, enhancer
and origin of replication, a DHFR gene and the SV40
5 terminator.

Example 3

10

Tissue Distribution

Human Multiple Tissue Northern Blots (MTN I, MTN II,
and MTN III; Clontech) were probed to determine the tissue
distribution of human ZSIG-9 expression. A 40 bp probe
15 (SEQ ID NO: 13) was used to probe the blots.
The 5' end of the probe was radioactively labeled using T4
polynucleotide kinase and forward reaction buffer (GIBCO
BRL, Gaithersburg, MD) according to the manufacturer's
specifications. The probe was purified using a NUCTRAP
20 push column (Stratagene, La Jolla, CA). ExpressHyb™
(Clontech) solution was used for prehybridization and as a
hybridizing solution for the Northern blots.
Hybridization took place overnight at 42°C using 2×10^6
cpm/ml of labeled probe. The blots were then washed at
25 55°C in 1X SSC, 0.1% SDS. A 1.2 kb transcript was
detected. The signal was strongest in heart, placenta,
liver and kidney. An intermediate signal was detected in
spleen, thymus, prostate, testis, ovary, small intestine,
colon, peripheral blood lymphocytes, thyroid, spinal cord.
30 Weak signal was detected in lymph node, trachea, adrenal
gland and bone marrow.

EXAMPLE 4

Chromosomal Assignment and Placement of Zsig9.

5 Zsig9 was mapped to chromosome 12 using the
commercially available "GeneBridge 4 Radiation Hybrid
Panel" (Research Genetics, Inc., Huntsville, AL). The
GeneBridge 4 Radiation Hybrid Panel contains PCRable DNAs
from each of 93 radiation hybrid clones, plus two control
10 DNAs (the HFL donor and the A23 recipient). A publicly
available WWW server ([http://www-genome.wi.mit.edu/cgi-
bin/contig/rhmapper.pl](http://www-genome.wi.mit.edu/cgi-bin/contig/rhmapper.pl)) allows mapping relative to the
Whitehead Institute/MIT Center for Genome Research's
radiation hybrid map of the human genome (the "WICGR"
15 radiation hybrid map) which was constructed with the
GeneBridge 4 Radiation Hybrid Panel.

For the mapping of Zsig9 with the "GeneBridge 4 RH
Panel", 20 μ l reactions were set up in a PCRable 96-well
20 microtiter plate (Stratagene, La Jolla, CA) and used in a
"RoboCycler Gradient 96" thermal cycler (Stratagene). Each
of the 95 PCR reactions consisted of 2 μ l 10X KlenTaq PCR
reaction buffer (CLONTECH Laboratories, Inc., Palo Alto,
CA), 1.6 μ l dNTPs mix (2.5 mM each, PERKIN-ELMER, Foster
25 City, CA), 1 μ l sense primer, SEQ ID NO: 14, 1 μ l
antisense primer, SEQ ID NO: 15, 2 μ l "RediLoad"
(Research Genetics, Inc., Huntsville, AL), 0.4 μ l 50X
Advantage KlenTaq Polymerase Mix (Clontech Laboratories,
Inc.), 25 ng of DNA from an individual hybrid clone or
30 control and x μ l ddH₂O for a total volume of 20 μ l. The
reactions were overlaid with an equal amount of mineral
oil and sealed. The PCR cycler conditions were as follows:
an initial 1 cycle 5 minute denaturation at 95°C, 35 cycles
of a 1 minute denaturation at 95°C, 1 minute annealing at
35 62°C and 1.5 minute extension at 72°C, followed by a final
1 cycle extension of 7 minutes at 72°C. The reactions were

separated by electrophoresis on a 3% NuSieve GTG agarose gel (FMC Bioproducts, Rockland, ME).

The results showed that Zsig9 maps 344.72 cR_3000
5 from the top of the human chromosome 12 linkage group on
the WICGR radiation hybrid map. Proximal and distal
framework markers were WI-6672 (D12S1410) and RP_L41_1,
respectively. The use of the surrounding markers positions
Zsig9 in the 12q15 region on the integrated LDB chromosome
10 12 map (The Genetic Location Database, University of
Southampton, WWW server: http://cedar.genetics.soton.ac.uk/public_html/) .

Example 5

15 Northern Blot Analysis of Human Tumors

Northern Analysis was carried out on the following
Tumor Blots ; Human Tumor Panel Blot I, Human Tumor Panel
Blot II, Human Tumor Panel Blot V, Human Stomach Tumor
20 Blot, and Human Colon Tumor Blot (Clontech, Palo Alto,
California). A probe was obtained using a PCR product
representing the full length coding sequence of zsig9.
The probe was radioactively labeled with 32P using
Rediprime Labeling System from Amersham (England). The
25 probe was purified using a NUCTRAP push column (Stratagene
Cloning Systems, La Jolla, Ca.) . EXPRESSHYB (Clontech,
Palo Alto, Ca.) solution was used for prehybridization and
hybridization. The hybridization solution consisted of 8
mls EXPRESSHYB, 80 µl Sheared Salmon Sperm DNA (10mg/ml, 5
30 Prime-3 Prime, Boulder, CO) , 48 µl Human Cot-1 DNA
(1mg/ml, Gibco BRL) and 20 µl labeled probe (8 x 10⁻⁵
CPM/µl). Hybridization took place overnight at 55°C And the
blots were then washed in 2X SSC, 0.1% SDS at RT, then 2X
SSC, 0.1% SDS at 60°C, followed by 0.1X SSC, 0.1% SDS wash
35 at 60°C. The blots were exposed overnight and developed.

A transcript of .8-1kb was observed in the following tissues. The strongest signals were in brain tumor, liver tumor, esophageal tumor, stomach tumor, colon tumor, 5 rectal tumor, and thyroid tumor. Weaker signals were in adrenal tumor and normal adrenal, peritoid tumor, and lymphoma tumor. Weakest signals were observed in normal liver, normal esophagus, normal stomach, normal colon, normal rectum, normal thyroid, and normal lymphoma. The 10 Stomach and Colon Tumor Blots showed signals consistent with those observed in the panel blots for stomach and colon tissue.

Example 6

15

Chromosomal Assignment and Placement of murine Zsig9

Murine Zsig9 was mapped in mouse to chromosome 10 using the commercially available mouse T31 whole genome 20 radiation hybrid (WGRH) panel (Research Genetics, Inc., Huntsville, AL) and the Map Manager QT linkage analysis program. At $P = 0.0001$, murine Zsig9 linked proximal to D10Mit136 at 62.0 cM with a LOD score of 4.3. This is a known region of synteny or linkage conservation with 25 regions of human chromosome 12.

The T31 WGRH panel contains PCRable DNAs from each of 100 radiation hybrid clones, plus two control DNAs (the 129aa donor and the A23 recipient). For the mapping of 30 murine Zsig9 with the T31 WGRH panel, 20 μ l reactions were set up in PCRable 96-well microtiter plates (Stratagene, La Jolla, CA) and used in "RoboCycler Gradient 96" thermal cyclers (Stratagene). Each of the 102 PCR reactions consisted of 2 μ l 10X KlenTaq PCR reaction buffer 35 (CLONTECH Laboratories, Inc., Palo Alto, CA), 1.6 μ l dNTPs mix (2.5 mM each, PERKIN-ELMER, Foster City, CA), 1 μ l

sense primer, (SEQ ID NO:22), 5' TCG CGC GAG AGT TTG GAG
3', 1 μ l antisense primer, (SEQ ID NO:23), 5' CCC AGC TTC
CCG CAC TTA 3', 2 μ l "RediLoad" (Research Genetics, Inc.,
Huntsville, AL), 0.4 μ l 50X Advantage KlenTaq Polymerase
5 Mix (Clontech Laboratories, Inc.), 25 ng of DNA from an
individual hybrid clone or control and x μ l ddH₂O for a
total volume of 20 μ l. The reactions were overlaid with an
equal amount of mineral oil and sealed. The PCR cycler
conditions were as follows: an initial 1 cycle 5 minute
10 denaturation at 94°C, 35 cycles of a 1 minute denaturation
at 94°C, 1 minute annealing at 62°C and 1.5 minute
extension at 72°C, followed by a final 1 cycle extension of
7 minutes at 72°C. The reactions were separated by
electrophoresis on a 2% agarose gel (Life Technologies,
15 Gaithersburg, MD). The use of the surrounding markers
positions murine Zsig9 in the 51-62 centiMorgan (cM)
region on mouse chromosome 10 map. Other genes of interest
which lie in or near this region are glioma-associated
oncogene homolog, myogenic factors 5 and 6, mast cell
20 growth factor, insulin-like growth factor, and tumor
rejection antigen-1.

CLAIMS

We claim:

1. An isolated polynucleotide which encodes a mammalian secretory-9 (Zsig9) polypeptide.
2. An isolated polynucleotide which encodes a mammalian Zsig9 polypeptide wherein said polynucleotide encodes a polypeptide selected from the group SEQ ID NOs:2-6, 17, 20 19 and 21 or a polypeptide which is at least 90% identical to the polypeptides of said group.
3. An isolated polynucleotide which encodes a peptide or polypeptide having at least 15 amino acid residues comprised of an epitope-bearing portion of a polypeptide of SEQ ID NOs: 2-6, 17, 20 19 and 21 or a polypeptide which is at least 90% identical to said polypeptides.
4. The polynucleotide of claim 3 wherein the peptide or polypeptide is fused to a carrier polypeptide or other carrier molecule.
5. An expression vector comprising the following operably linked elements:
 - a transcription promoter;
 - a DNA segment which encodes a Zsig9 polypeptide or a peptide or polypeptide which contains an epitope-bearing region of a Zsig9 polypeptide; and
 - a transcription terminator.
6. An expression vector comprising the following operably linked elements:
 - (a) a transcription promoter;
 - (b) a DNA segment encoding a chimeric polypeptide, wherein said chimeric polypeptide consists essentially of a first portion and a second portion joined by a peptide bond,

said first portion being comprised of a mammalian polypeptide, said polypeptide being the amino acid sequences of SEQ ID NOs: 2-6, 17, 20 19 and 21 and said second portion being a second polypeptide or protein.

(c) a transcription terminator.

7. An isolated mammalian Zsig9 polypeptide

8. An isolated Zsig9 polypeptide selected from the group of amino acid sequences consisting of SEQ ID NOs: 22-6, 17, 20 19 and 21 or a polypeptide which is at least 90% identical to said polypeptides.

9. An isolated peptide or polypeptide having at least 15 amino acid residues comprised of an epitope-bearing portion of a polypeptide of SEQ ID NOs: 2-6, 17, 20 19 and 21.

10. An antibody, antibody fragment or single-chain antibody that specifically binds to a mammalian polypeptide, said polypeptide being defined by the amino acid sequences of SEQ ID NOs: 2-6, 17, 20 19 and 21.

11. An antibody of claim 10 wherein said antibody is either monoclonal or polyclonal.

12. The antibody, antibody fragment or single-chain antibody of claim 10 wherein said antibody, antibody fragment or single-chain antibody is humanized.

13. A method for producing an antibody which binds to a mammalian Zsig9 polypeptide or a peptide or polypeptide defined by SEQ ID NOs: 2-6, 17, 20 19 and 21 or to a peptide or polypeptide which is at least 90% identical to said peptide or polypeptide comprising bringing into contact a mammalian Zsig9 polypeptide or a peptide or polypeptide defined by SEQ ID NOs: 2-6, 17, 20 19 and 21 or to a peptide or polypeptide

which is at least 90% identical to said peptide or polypeptide with a cell capable of producing antibodies or the cell is brought into contact with a nucleic acid which encodes said peptide or polypeptide, wherein said cell produces antibodies to said peptide or polypeptide; and
isolating said antibody.

14. The method of claim 13 wherein an animal is inoculated with the peptide or polypeptide or nucleic acid under conditions wherein the animal produces antibodies to said peptide; and isolating said antibodies

15. The method of claims 13 or 14 wherein the antibodies are polyclonal or monoclonal.

16. An anti-idiotypic antibody, anti-idiotypic antibody fragment or anti-idiotypic single-chain antibody which binds to an antibody, an antibody fragment or single-chain antibody of peptide or polypeptide defined by SEQ ID NOS: 2-6, 17, 20 19 and 21 or to a peptide or polypeptide which is at least 90% identical to said peptide or polypeptide

SEQUENCE LISTING

(1) GENERAL INFORMATION

- (i) APPLICANT: ZymoGenetics, Inc.
1201 Eastlake Avenue East
Seattle
WA
USA
98102
- (ii) TITLE OF THE INVENTION: MAMMALIAN ZSIG9
- (iii) NUMBER OF SEQUENCES: 24
- (iv) CORRESPONDENCE ADDRESS:
 - (A) ADDRESSEE: ZymoGenetics Inc.
 - (B) STREET: 1201 Eastlake Ave East
 - (C) CITY: Seattle
 - (D) STATE: WA
 - (E) COUNTRY: USA
 - (F) ZIP: 98102
- (v) COMPUTER READABLE FORM:
 - (A) MEDIUM TYPE: Diskette
 - (B) COMPUTER: IBM Compatible
 - (C) OPERATING SYSTEM: DOS
 - (D) SOFTWARE: FastSEQ for Windows Version 2.0
- (vi) CURRENT APPLICATION DATA:
 - (A) APPLICATION NUMBER:
 - (B) FILING DATE:
 - (C) CLASSIFICATION:
- (vii) PRIOR APPLICATION DATA:
 - (A) APPLICATION NUMBER:
 - (B) FILING DATE:
- (viii) ATTORNEY/AGENT INFORMATION:
 - (A) NAME: Lunn, Paul G

(B) REGISTRATION NUMBER: 32,743
 (C) REFERENCE/DOCKET NUMBER: 97-11PC

(ix) TELECOMMUNICATION INFORMATION:

(A) TELEPHONE: 206-442-6627
 (B) TELEFAX: 206-442-6678
 (C) TELEX:

(2) INFORMATION FOR SEQ ID NO:1:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 649 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: double
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(ix) FEATURE:

(A) NAME/KEY: Coding Sequence
 (B) LOCATION: 104...354
 (D) OTHER INFORMATION:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:

CGGCCCAAGG	CTGGGGCCAA	AGTGAAAGTC	CAGCGGTCTN	CCAGCGCTTG	GGCCACGGCG	60
GCGGCCCTGG	GACCAAAGGT	GGAGCAACCC	CGTTACCCTA	AAR ATG AAA	GGC TGG	115
				Met Lys Gly Trp		
				1		
GGT TGG CTG GCC CTG CTT CTG GGG GCC CTG CTG GGA ACC GCC TGG GCT						163
Gly Trp Leu Ala Leu Leu Leu Gly Ala Leu Leu Gly Thr Ala Trp Ala						
5	10		15		20	
CGG AGG AGC CAG GAT CTC CAC TGT GGA GCA TGC AGG GCT CTG GTG GAT						211
Arg Arg Ser Gln Asp Leu His Cys Gly Ala Cys Arg Ala Leu Val Asp						
	25		30		35	
GAA CTA GAA TGG GAA ATT GCC CAG GTG GAC CCC AAG AAG ACC ATT CAG						259
Glu Leu Glu Trp Glu Ile Ala Gln Val Asp Pro Lys Lys Thr Ile Gln						
	40		45		50	

ATG GGA TCT TTC CGG ATC AAT CCA GAT GGC AGC CAG TCA GTG GTG GAG	307
Met Gly Ser Phe Arg Ile Asn Pro Asp Gly Ser Gln Ser Val Val Glu	
55 60 65	
GTA ACT GTT ACT GTT CCC CCA AAC AAA GTA GCT CAC TCT GGC TTT GG AT	356
Val Thr Val Thr Val Pro Pro Asn Lys Val Ala His Ser Gly Phe Gly	
70 75 80	
GAAATTCGAT TGCTTAAAAA GGACCTTGGT TTAATAGAAA TGAAGAAAAC AGACTCAGAA	416
AAAAGATTG GCTCTGTCTC ATTTGGAAGA AGCTGCAGGC TTATTCCCA TGCACCTTGCT	476
TCCTGGCTGC AAACCTTAAT ACTTTGTTN TGCTGTAGAA TTTGTTAGCA AACAGGGAGT	536
CCTGATCAGC ACCCTTNTCC ACATCCACAT GACTGGTTTT TAATGTAGCA CTGTGGTATA	596
CATGCAAACA TTCCGTTCAA AATCTGAGTC GGAGCTAAAA AAAAAAAAAA AAA	649

(2) INFORMATION FOR SEQ ID NO:2:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 84 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(v) FRAGMENT TYPE: internal

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

Met	Lys	Gly	Trp	Gly	Trp	Leu	Ala	Leu	Leu	Gly	Ala	Leu	Leu	Gly
1				5				10				15		
Thr	Ala	Trp	Ala	Arg	Arg	Ser	Gln	Asp	Leu	His	Cys	Gly	Ala	Cys
			20					25				30		Arg
Ala	Leu	Val	Asp	Glu	Leu	Glu	Trp	Glu	Ile	Ala	Gln	Val	Asp	Pro
			35					40				45		Lys
Lys	Thr	Ile	Gln	Met	Gly	Ser	Phe	Arg	Ile	Asn	Pro	Asp	Gly	Ser
	50					55					60			Gln
Ser	Val	Val	Glu	Val	Thr	Val	Thr	Val	Pro	Pro	Asn	Lys	Val	Ala
65					70					75				80
Ser	Gly	Phe	Gly											

(2) INFORMATION FOR SEQ ID NO:3:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 64 amino acids
- (B) TYPE: amino acid

(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:

Arg	Arg	Ser	Gln	Asp	Leu	His	Cys	Gly	Ala	Cys	Arg	Ala	Leu	Val	Asp
1				5				10					15		
Glu	Leu	Glu	Trp	Glu	Ile	Ala	Gln	Val	Asp	Pro	Lys	Lys	Thr	Ile	Gln
			20					25					30		
Met	Gly	Ser	Phe	Arg	Ile	Asn	Pro	Asp	Gly	Ser	Gln	Ser	Val	Val	Glu
			35				40					45			
Val	Thr	Val	Thr	Val	Pro	Pro	Asn	Lys	Val	Ala	His	Ser	Gly	Phe	Gly
			50				55				60				

(2) INFORMATION FOR SEQ ID NO:4:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 62 amino acids
(B) TYPE: amino acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:

Ser	Gln	Asp	Leu	His	Cys	Gly	Ala	Cys	Arg	Ala	Leu	Val	Asp	Glu	Leu
1				5				10					15		
Glu	Trp	Glu	Ile	Ala	Gln	Val	Asp	Pro	Lys	Lys	Thr	Ile	Gln	Met	Gly
			20					25					30		
Ser	Phe	Arg	Ile	Asn	Pro	Asp	Gly	Ser	Gln	Ser	Val	Val	Glu	Val	Thr
			35				40					45			
Val	Thr	Val	Pro	Pro	Asn	Lys	Val	Ala	His	Ser	Gly	Phe	Gly		
			50				55				60				

(2) INFORMATION FOR SEQ ID NO:5:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 25 amino acids
(B) TYPE: amino acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:5:

Ser Gln Asp Leu His Cys Gly Ala Cys Arg Ala Leu Val Asp Glu Leu
 1 5 10 15
 Glu Trp Glu Ile Ala Gln Val Asp Pro
 20 25

(2) INFORMATION FOR SEQ ID NO:6:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 35 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:6:

Thr Ile Gln Met Gly Ser Phe Arg Ile Asn Pro Asp Gly Ser Gln Ser
 1 5 10 15
 Val Val Glu Val Thr Val Thr Val Pro Pro Asn Lys Val Ala His Ser
 20 25 30
 Gly Phe Gly
 35

(2) INFORMATION FOR SEQ ID NO:7:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 417 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(vi) ORIGINAL SOURCE:

- (A) ORGANISM: EST192508

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:7:

CTGGGGCAAA	GTGAGAGTCC	AGCGGTCTTC	CAGCGCTTGG	GCCACGGCGG	CGGCCTGGGA	60
GCAGAGGTGG	AGCGACCCCA	TTACGCTAAA	GATGAAAGGC	TGGGGTTGGC	TGGCCCTGCT	120
TCTGGGGGCC	CTGCTGGGAA	CCGCCTGGGC	TCGGAGGAGC	AGGGATCTCC	ACTGTGGAGC	180
ATGCAGGGCT	CTGGTGGATG	AACTAGAATG	GGAAATTGCC	CAGGTGGACC	CCAAGAAGAC	240
CATTCAGATG	GGATCTTTCC	GGATCAATCC	AGATGGCAGC	CAGTCAGTGG	TNGAGGTAAC	300

TGTTACTGTT CCCCCAAACA AAGTAGCTCA CTCTGGCTTT NGATGAATTT CGATTNTTT 360
 AAAAAGGACC TTTGTTTTAT TAGGAATTGA AGAAAACAGA TTCAGAAAAA AGTTT
 417.

(2) INFORMATION FOR SEQ ID NO:8:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 10 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(vi) ORIGINAL SOURCE:

- (A) ORGANISM: Flag

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:8:

Asp	Tyr	Lys	Asp	Asp	Asp	Asp	Lys	Gly	Ser
1				5					10

(2) INFORMATION FOR SEQ ID NO:9:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 25 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(vi) ORIGINAL SOURCE:

- (A) ORGANISM: zc13654

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:9:

GCGCGGATCC CGGAGGAGCC AGGAT 25

(2) INFORMATION FOR SEQ ID NO:10:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 25 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(iv) ANTISENSE: YES

(vi) ORIGINAL SOURCE:
(A) ORGANISM: ZC13,655

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:10:

CGCGCTCGAG TCATCCAAAG CCAGA

25

(2) INFORMATION FOR SEQ ID NO:11:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 25 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: Other

(vi) ORIGINAL SOURCE:
(A) ORGANISM: ZC13656

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:11:

GCGCGAATTC ATGAAAGGCT GGGGT

25

(2) INFORMATION FOR SEQ ID NO:12:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 25 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(iv) ANTISENSE: YES

(vi) ORIGINAL SOURCE:
(A) ORGANISM: ZC13657

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:12:

CGCGGGATCC TCCAAAGCCA GAGTG

25

(2) INFORMATION FOR SEQ ID NO:13:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 40 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(iv) ANTISENSE: YES

(vi) ORIGINAL SOURCE:

(A) ORGANISM: ZC11.666

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:13:

TTCATCCACC AGAGCCCTGC ATGCTCCACA GTGGAGATCC

40

(2) INFORMATION FOR SEQ ID NO:14:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 18 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(vi) ORIGINAL SOURCE:

(A) ORGANISM: zc 14,477

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:14:

GGGCTCTGGT GGATGAAC

18

(2) INFORMATION FOR SEQ ID NO:15:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 18 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(iv) ANTISENSE: YES

(vi) ORIGINAL SOURCE:

(A) ORGANISM: ZC 14,489

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:15:

TACCTCCACC ACTGACTG

18

(2) INFORMATION FOR SEQ ID NO:16:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 806 base pairs

(B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(ix) FEATURE:

(A) NAME/KEY: Coding Sequence

(B) LOCATION: 104...649

(D) OTHER INFORMATION:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:16:

CGGCCCAAGG CTGGGGCCAA AGTGAAAGTC CAGCGGTCTN CCAGCGCTTG GGCCACGGCG	60
GCGGCCCTGG GACCAAAGGT GGAGCAACCC CGTTACCCTA AAR ATG AAA GGC TGG	115
Met Lys Gly Trp	
1	
GGT TGG CTG GCC CTG CTT CTG GGG GCC CTG CTG GGA ACC GCC TGG GCT	163
Gly Trp Leu Ala Leu Leu Leu Gly Ala Leu Leu Gly Thr Ala Trp Ala	
5 10 15 20	
CGG AGG AGC CAG GAT CTC CAC TGT GGA GCA TGC AGG GCT CTG GTG GAT	211
Arg Arg Ser Gln Asp Leu His Cys Gly Ala Cys Arg Ala Leu Val Asp	
25 30 35	
GAA CTA GAA TGG GAA ATT GCC CAG GTG GAC CCC AAG AAG ACC ATT CAG	259
Glu Leu Glu Trp Glu Ile Ala Gln Val Asp Pro Lys Lys Thr Ile Gln	
40 45 50	
ATG GGA TCT TTC CGG ATC AAT CCA GAT GGC AGC CAG TCA GTG GTG GAG	307
Met Gly Ser Phe Arg Ile Asn Pro Asp Gly Ser Gln Ser Val Val Glu	
55 60 65	
GTG CCT TAT GCC CGC TCA GAG GCC CAC CTC ACA GAG CTG CTG GAG GAG	355
Val Pro Tyr Ala Arg Ser Glu Ala His Leu Thr Glu Leu Leu Glu Glu	
70 75 80	
ATA TGT GAC CGG ATG AAG GAG TAT GGG GAA CAG ATT GAT CCT TCC ACC	403
Ile Cys Asp Arg Met Lys Glu Tyr Gly Glu Gln Ile Asp Pro Ser Thr	
85 90 95 100	
CAT CGC AAG AAC TAC GTA CGT GTA GTG GGC CGG AAT GGA GAA TCC AGT	451
His Arg Lys Asn Tyr Val Arg Val Val Gly Arg Asn Gly Glu Ser Ser	
105 110 115	

GAA CTG GAC CTA CAA GGC ATC CGA ATC GAC TCA GAT ATT AGC GGC ACC	499
Glu Leu Asp Leu Gln Gly Ile Arg Ile Asp Ser Asp Ile Ser Gly Thr	
120 125 130	
CTC AAG TTT GCG TGT GAG AGC ATT GTG GAG GAA TAC GAG GAT GAA CTC	547
Leu Lys Phe Ala Cys Glu Ser Ile Val Glu Glu Tyr Glu Asp Glu Leu	
135 140 145	
ATT GAA TTC TTT TCC CGA GAG GCT GAC AAT GTT AAA GAC AAA CTT TGC	595
Ile Glu Phe Phe Ser Arg Glu Ala Asp Asn Val Lys Asp Lys Leu Cys	
150 155 160	
AGT AAG CGA ACA GAT CTT TGT GAC CAT GCC CTG CAC ATA TCG CAT GAT	643
Ser Lys Arg Thr Asp Leu Cys Asp His Ala Leu His Ile Ser His Asp	
165 170 175 180	
GAG CTA TGAACCACTG GAGCAGCCCA CACTGGCTTG ATGGATCACC CCCAGGAGGG GA	701
Glu Leu	
AAATGGTGGC AATGCCTTTT ATATATTATG TTTTACTGA AATTAAGTGA AAAAAATATGA	761
AACCAAAAGT AAAAAAAAAA AAAAAAGAG AGAGAGAGAG AACTA	806

(2) INFORMATION FOR SEQ ID NO:17:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 182 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(v) FRAGMENT TYPE: internal

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:17:

Met	Lys	Gly	Trp	Gly	Trp	Leu	Ala	Leu	Leu	Leu	Gly	Ala	Leu	Leu	Gly
1				5				10					15		
Thr	Ala	Trp	Ala	Arg	Arg	Ser	Gln	Asp	Leu	His	Cys	Gly	Ala	Cys	Arg
			20				25					30			
Ala	Leu	Val	Asp	Glu	Leu	Glu	Trp	Glu	Ile	Ala	Gln	Val	Asp	Pro	Lys
		35				40					45				
Lys	Thr	Ile	Gln	Met	Gly	Ser	Phe	Arg	Ile	Asn	Pro	Asp	Gly	Ser	Gln
50					55					60					

11

Ser Val Val Glu Val Pro Tyr Ala Arg Ser Glu Ala His Leu Thr Glu
 65 70 75 80
 Leu Leu Glu Glu Ile Cys Asp Arg Met Lys Glu Tyr Gly Glu Gln Ile
 85 90 95
 Asp Pro Ser Thr His Arg Lys Asn Tyr Val Arg Val Val Gly Arg Asn
 100 105 110
 Gly Glu Ser Ser Glu Leu Asp Leu Gln Gly Ile Arg Ile Asp Ser Asp
 115 120 125
 Ile Ser Gly Thr Leu Lys Phe Ala Cys Glu Ser Ile Val Glu Glu Tyr
 130 135 140
 Glu Asp Glu Leu Ile Glu Phe Phe Ser Arg Glu Ala Asp Asn Val Lys
 145 150 155 160
 Asp Lys Leu Cys Ser Lys Arg Thr Asp Leu Cys Asp His Ala Leu His
 165 170 175
 Ile Ser His Asp Glu Leu
 180

(2) INFORMATION FOR SEQ ID NO:18:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 1069 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(ix) FEATURE:

- (A) NAME/KEY: Coding Sequence
- (B) LOCATION: 358...903
- (D) OTHER INFORMATION:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:18:

GAATTCGGCA CGAGGGGGGT CCTCGCTGCC TCGGAGGCGC TCCTAAAGCT GCCTGCTCGC 60
 GCGAGAGTTT GGAGGGGCGG GCTTAGGGTC AGTTTCGGTG GGGGGCTCGC ACGGGACCCT 120
 CAGATCTCCG CTTAGGTGCC TAGTTAAGTG CGGGAAGCTG GGCCAGGCGG TCACTGGCCA 180
 CCCTGAACCT GGCGGGAGCC GGAGCGCTCT GGAGAAGCCG GGACAGCCCC GTTTTTCCCA 240
 GCCAGCTGCT AGGGTTGGGA CCCACAGAAA ACAAAAGTGAG AGTCCGGCTG CTTTCCAGAG 300
 CCTGGGCCAC GGCGGCGGCC GTGGGAGCAG AGGTGGAGCG ACCCTGTTAC ACTAAAG ATG 360
 Met
 1

AAA GGC TGG GGT TGG CTA GCC CTA CTT TTG GGG GTC CTG CTG GGA ACT	408
Lys Gly Trp Gly Trp Leu Ala Leu Leu Leu Gly Val Leu Leu Gly Thr	
5 10 15	
GCC TGG GCT CGA AGG AGC CAA GAT CTA CAC TGT GGA GCT TGC AGG GCT	456
Ala Trp Ala Arg Arg Ser Gln Asp Leu His Cys Gly Ala Cys Arg Ala	
20 25 30	
CTG GTG GAT GAA TTA GAG TGG GAA ATT GCC CGC GTG GAC CCC AAG AAG	504
Leu Val Asp Glu Leu Glu Trp Glu Ile Ala Arg Val Asp Pro Lys Lys	
35 40 45	
ACC ATT CAG ATG GGA TCC TTC CGA ATC AAT CCA GAT GGC AGC CAG TCA	552
Thr Ile Gln Met Gly Ser Phe Arg Ile Asn Pro Asp Gly Ser Gln Ser	
50 55 60 65	
GTT GTG GAG GTA CCT TAT GCC CGC TCA GAG GCC CAC CTC ACA GAG TTG	600
Val Val Glu Val Pro Tyr Ala Arg Ser Glu Ala His Leu Thr Glu Leu	
70 75 80	
CTT GAG GAG GTG TGT GAC CGA ATG AAG GAG TAC GGG GAA CAG ATT GAC	648
Leu Glu Glu Val Cys Asp Arg Met Lys Glu Tyr Gly Glu Gln Ile Asp	
85 90 95	
CCT TCT ACC CAC CGC AAG AAC TAC GTA CGC GTC GTG AGC CGG AAT GGA	696
Pro Ser Thr His Arg Lys Asn Tyr Val Arg Val Val Ser Arg Asn Gly	
100 105 110	
GAA TCC AGT GAA CTA GAC TTA CAG GGC ATC CGA ATT GAC TCA GAT ATC	744
Glu Ser Ser Glu Leu Asp Leu Gln Gly Ile Arg Ile Asp Ser Asp Ile	
115 120 125	
AGC GGC ACC CTC AAG TTT GCG TGT GAG AGC ATT GTG GAA GAA TAC GAG	792
Ser Gly Thr Leu Lys Phe Ala Cys Glu Ser Ile Val Glu Glu Tyr Glu	
130 135 140 145	
GAT GAG CTT ATC GAA TTC TTC TCC AGA GAG GCT GAC AAC GTT AAA GAC	840
Asp Glu Leu Ile Glu Phe Phe Ser Arg Glu Ala Asp Asn Val Lys Asp	
150 155 160	
AAA CTT TGC AGT AAG CGG ACA GAT CTA TGT GAC CAT GCC CTG CAC AGA	888
Lys Leu Cys Ser Lys Arg Thr Asp Leu Cys Asp His Ala Leu His Arg	
165 170 175	

(2) INFORMATION FOR SEQ ID NO:19:

(A) LENGTH: 182 amino acids
(B) TYPE: amino acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(v) FRAGMENT TYPE: internal

[illegible]

(2) INFORMATION FOR SEQ ID NO:20:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 162 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:20:

```

Arg Arg Ser Gln Asp Leu His Cys Gly Ala Cys Arg Ala Leu Val Asp
 1           5           10          15
Glu Leu Glu Trp Glu Ile Ala Gln Val Asp Pro Lys Lys Thr Ile Gln
 20          25          30
Met Gly Ser Phe Arg Ile Asn Pro Asp Gly Ser Gln Ser Val Val Glu
 35          40          45
Val Pro Tyr Ala Arg Ser Glu Ala His Leu Thr Glu Leu Leu Glu Glu
 50          55          60
Ile Cys Asp Arg Met Lys Glu Tyr Gly Glu Gln Ile Asp Pro Ser Thr
 65          70          75          80
His Arg Lys Asn Tyr Val Arg Val Val Gly Arg Asn Gly Glu Ser Ser
 85          90          95
Glu Leu Asp Leu Gln Gly Ile Arg Ile Asp Ser Asp Ile Ser Gly Thr
100         105         110
Leu Lys Phe Ala Cys Glu Ser Ile Val Glu Glu Tyr Glu Asp Glu Leu
115         120         125
Ile Glu Phe Phe Ser Arg Glu Ala Asp Asn Val Lys Asp Lys Leu Cys
130         135         140
Ser Lys Arg Thr Asp Leu Cys Asp His Ala Leu His Ile Ser His Asp
145         150         155         160
Glu Leu

```

(2) INFORMATION FOR SEQ ID NO:21:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 162 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

15

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:21:

Arg	Arg	Ser	Gln	Asp	Leu	His	Cys	Gly	Ala	Cys	Arg	Ala	Leu	Val	Asp
1			5					10					15		
Glu	Leu	Glu	Trp	Glu	Ile	Ala	Arg	Val	Asp	Pro	Lys	Lys	Thr	Ile	Gln
		20						25					30		
Met	Gly	Ser	Phe	Arg	Ile	Asn	Pro	Asp	Gly	Ser	Gln	Ser	Val	Val	Glu
		35					40					45			
Val	Pro	Tyr	Ala	Arg	Ser	Glu	Ala	His	Leu	Thr	Glu	Leu	Leu	Glu	Glu
	50					55				60					
Val	Cys	Asp	Arg	Met	Lys	Glu	Tyr	Gly	Glu	Gln	Ile	Asp	Pro	Ser	Thr
65					70				75					80	
His	Arg	Lys	Asn	Tyr	Val	Arg	Val	Val	Ser	Arg	Asn	Gly	Glu	Ser	Ser
			85					90					95		
Glu	Leu	Asp	Leu	Gln	Gly	Ile	Arg	Ile	Asp	Ser	Asp	Ile	Ser	Gly	Thr
		100					105					110			
Leu	Lys	Phe	Ala	Cys	Glu	Ser	Ile	Val	Glu	Glu	Tyr	Glu	Asp	Glu	Leu
	115						120					125			
Ile	Glu	Phe	Phe	Ser	Arg	Glu	Ala	Asp	Asn	Val	Lys	Asp	Lys	Leu	Cys
	130					135					140				
Ser	Lys	Arg	Thr	Asp	Leu	Cys	Asp	His	Ala	Leu	His	Arg	Ser	His	Asp
145					150					155				160	
Glu	Leu														

(2) INFORMATION FOR SEQ ID NO:22:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 18 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: Other

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:22:

TCGCGCGAGA GTTTGGAG

18

(2) INFORMATION FOR SEQ ID NO:23:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 18 base pairs
- (B) TYPE: nucleic acid

16

(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: Other

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:23:

CCCAGCTTCC CGCACTTA

18

(2) INFORMATION FOR SEQ ID NO:24:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 35 amino acids
(B) TYPE: amino acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:24:

[illegible]

INTERNATIONAL SEARCH REPORT

International Application No

PCT/US 98/13859

A. CLASSIFICATION OF SUBJECT MATTER

IPC 6 C12N15/12 C12N15/62 C07K14/47 C07K16/18 C07K16/42
A61K38/17

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC 6 C12N C07K A61K

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	DATABASE GENBANK Accession No. R22491, HILLIER L. ET AL.: "H. sapiens clone IMAGE 130610" XP002083365 cited in the application compare with Seq. ID 7.	1-9
A	---	10-16
A	WO 93 03157 A (DANA FARBER CANCER INST INC) 18 February 1993 see abstract --- -/--	6

☒ Further documents are listed in the continuation of box C.

☒ Patent family members are listed in annex.

* Special categories of cited documents :

"A" document defining the general state of the art which is not considered to be of particular relevance

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"O" document referring to an oral disclosure, use, exhibition or other means

"P" document published prior to the international filing date but later than the priority date claimed

"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention

"X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone

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"&" document member of the same patent family

Date of the actual completion of the international search

6 November 1998

Date of mailing of the international search report

23/11/1998

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Galli, I

INTERNATIONAL SEARCH REPORT

International Application No
PCT/US 98/13859

C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category	Citation of document, with indication where appropriate, of the relevant passages	Relevant to claim No.
A	<p>RATTNER A. ET AL.: "A family of secreted proteins contains homology to the cysteine-rich ligand-binding domain of frizzled receptors"</p> <p>PROC. NATL. ACAD. SCI. USA, vol. 94, April 1997, pages 2859-2863, XP002083363 see abstract see figure 2</p> <p>-----</p>	1-16

INTERNATIONAL SEARCH REPORT

information on patent family members

International Application No

PCT/US 98/13859

Patent document cited in search report	Publication date	Patent family member(s)	Publication date
WO 9303157	A	18-02-1993	NONE